

Introduction to R software

Correction of exercises

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Exercise 1

```
# Number of flowers from setosa species
```

```
> sum(iris$Species=="setosa")
```

```
[1] 50
```

```
# Sepal length in increasing order
```

```
> sort(iris$Sepal.Length)
```

```
# Mean sepal length
```

```
> mean(iris$Sepal.Length)
```

```
[1] 5.843333
```

```
# Number of flowers with a sepal length higher than 5 cm
```

```
> sum(iris$Sepal.Length > 5 )
```

```
[1] 118
```

```
# Number of setosa flowers with a sepal length larger than 5 cm
```

```
> sum( (iris$Sepal.Length> 5) & (iris$Species=="setosa" ) )
```

```
[1] 22
```

Exercise 2

```
> sl = iris$Sepal.Length
```

```
# Class of sl object
```

```
> class(sl)
```

```
[1] "numeric"
```

```
# Minimal, maximal and mean sepal length
```

```
> min(sl)
```

```
[1] 4.3
```

```
> max(sl)
```

```
[1] 7.9
```

```
# Length of the 10 largest sepals
```

```
> sort(sl, decreasing=TRUE)[1:10]
```

```
[1] 7.9 7.7 7.7 7.7 7.7 7.6 7.4 7.3 7.2 7.2
```

Exercise 3

```
# Class of the Species object from the iris dataset
```

```
> class(iris$Species)
```

```
[1] "factor"
```

```
# Levels of this factor
```

```
> levels(iris$Species)
```

```
[1] "setosa" "versicolor" "virginica"
```

```
# Number of flowers for each species
```

```
> table(iris$Species)
```

```
setosa versicolor virginica
```

```
50      50      50
```

```
# Species of the flower with the smallest sepal length
```

```
> iris$Species[iris$Sepal.Length == min(iris$Sepal.Length)]
```

```
[1] setosa
```

Exercise 4

```
# Importation of the humanGenomeSummary.txt file into R
huge=read.table("humanGenomeSummary.txt", header=TRUE ,sep="\t")
```

```
# Visualisation of the created object
head(huge)
```

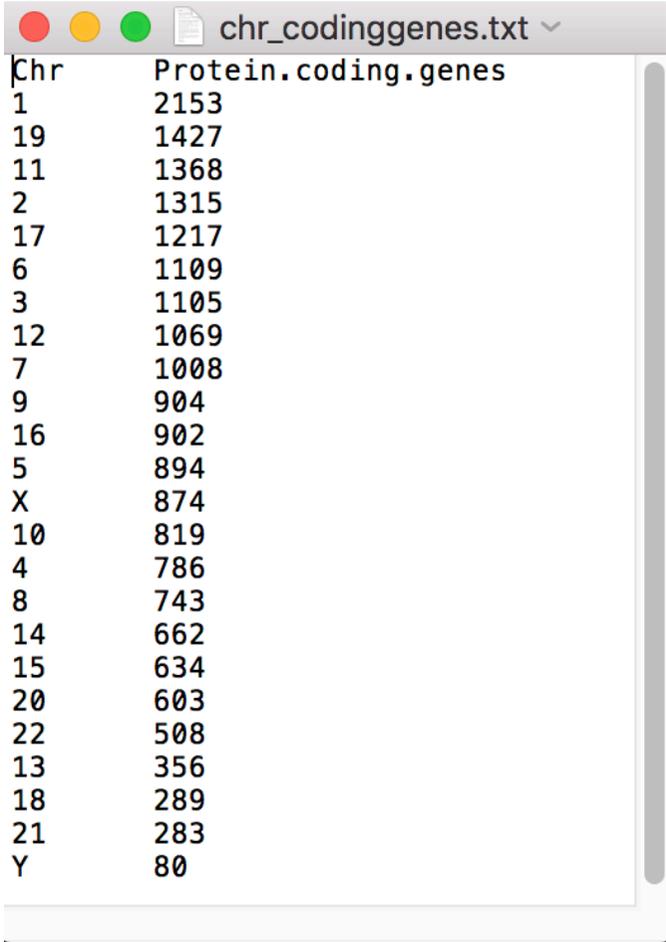
	Chr	Length	Protein.coding.genes	Pseudogenes	SNPs
1	1	247249719	2153	114	1059468
2	2	242951149	1315	88	1005820
3	3	199501827	1105	89	830895
4	4	191273063	786	74	871520
5	5	180857866	894	89	745224
6	6	170899992	1109	90	797289

```
# Total number of protein coding genes
sum(huge$Protein.coding.genes)
[1] 21108
```

```
# Chromosomes with more than 1,000 annotated protein coding genes
huge$Chr[huge$Protein.coding.genes>1000]
[1] 1 2 3 6 7 11 12 17 19
```

Exercise 4

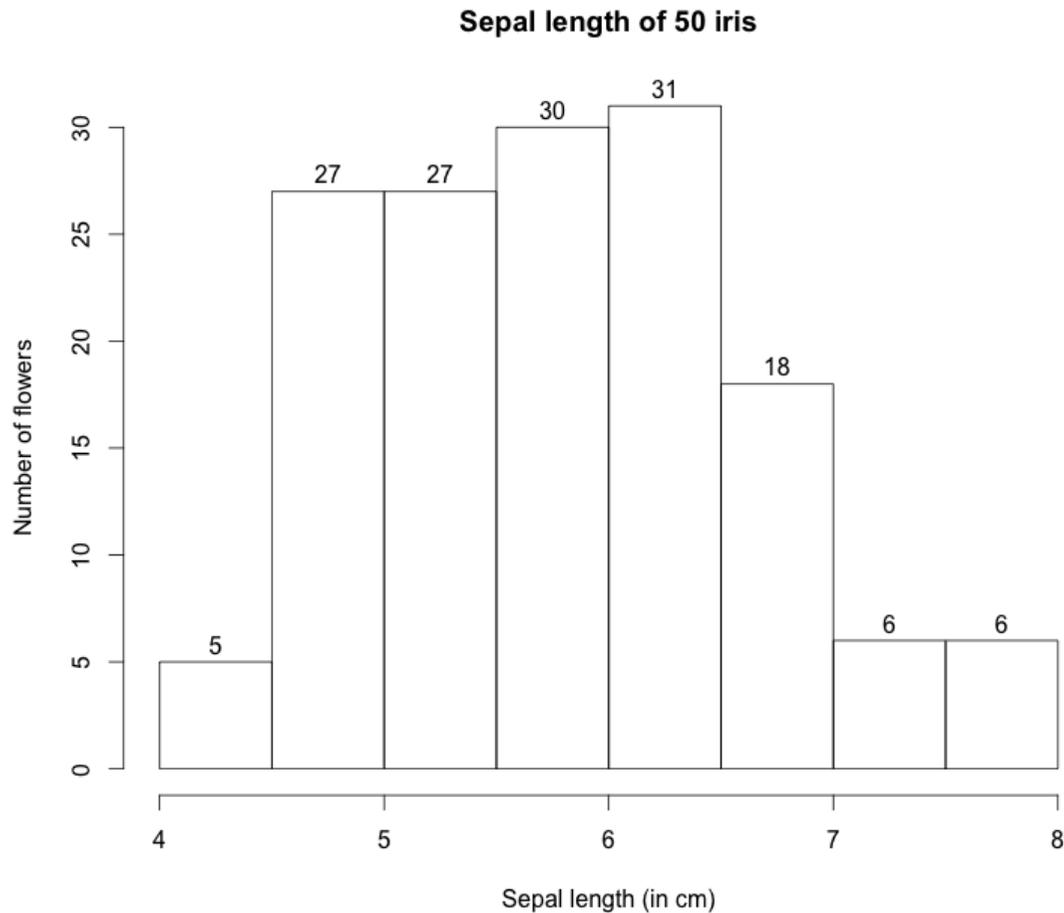
```
write.table( huge[order(huge$Protein.coding.genes, decreasing=TRUE), c(1,3)],  
            file="chr_codinggenes.txt", sep="\t", quote=FALSE, row.names=FALSE)
```



Chr	Protein.coding.genes
1	2153
19	1427
11	1368
2	1315
17	1217
6	1109
3	1105
12	1069
7	1008
9	904
16	902
5	894
X	874
10	819
4	786
8	743
14	662
15	634
20	603
22	508
13	356
18	289
21	283
Y	80

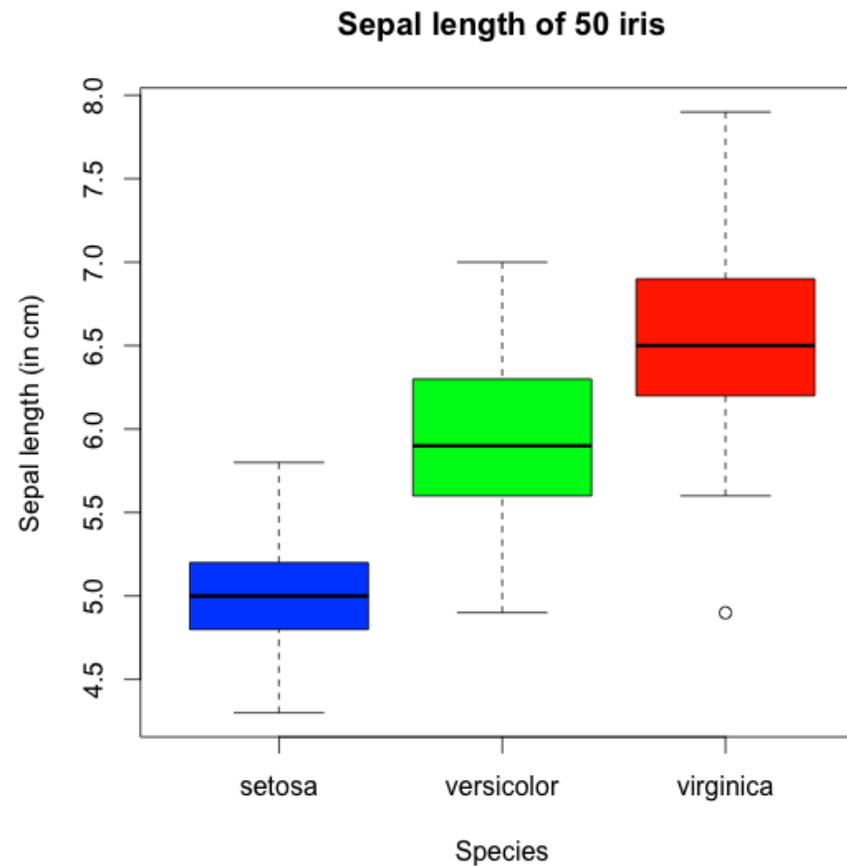
Exercise 5 : histogram

```
hist(iris$Sepal.Length, labels=TRUE, xlab="Sepal length (in cm)",  
ylab="Number of flowers", main="Sepal length of 50 iris")
```



Exercise 5 : boxplot

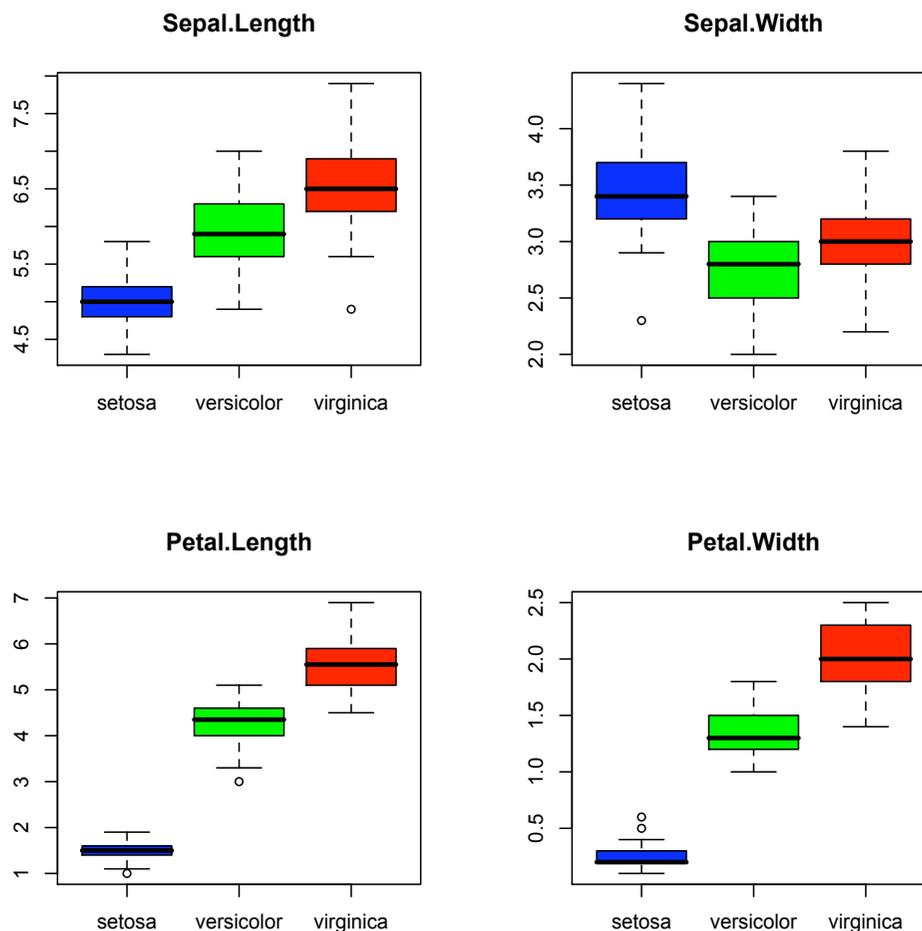
```
boxplot(iris$Sepal.Length ~ iris$Species, col=c("blue","green","red"),  
main= "Sepal length of 50 iris",  
xlab= "Species", ylab= "Sepal length (in cm)")
```



Exercise 5 :

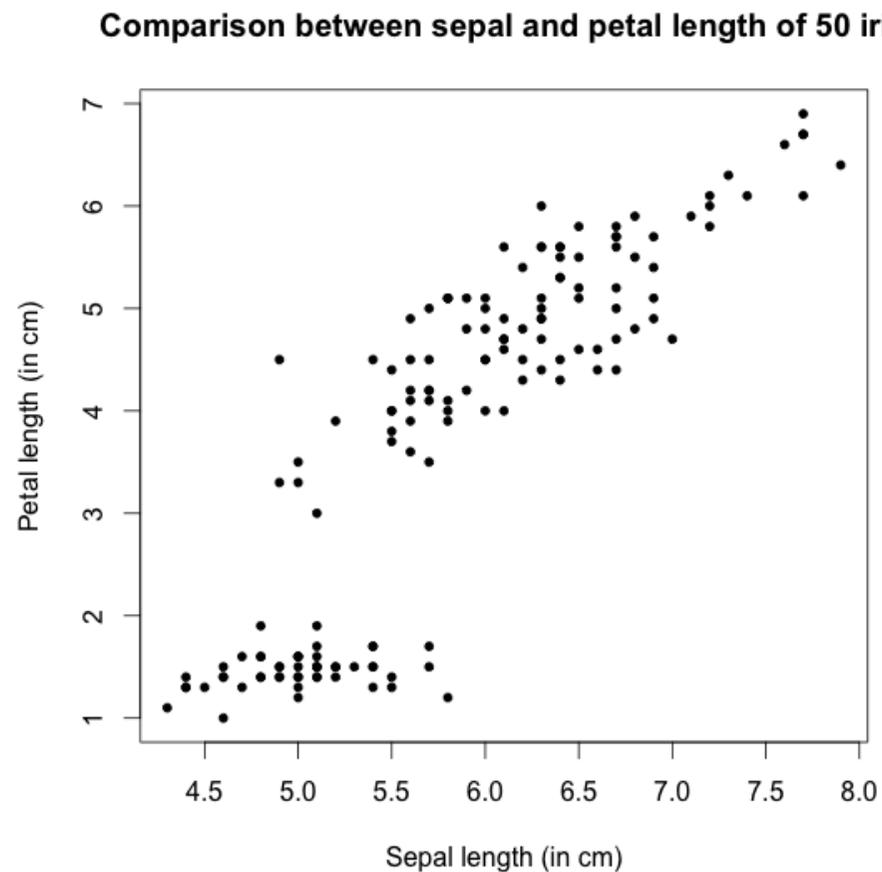
boxplots and graphics windows partition

```
par(mfrow=c(2,2))
for (i in 1:4){ # for each quantitative variable
  boxplot( iris[,i]~iris$Species, main=colnames(iris)[i], col=c("blue","green","red"))
}
```



Exercise 5 : scatterplot

```
plot(iris$Sepal.Length, iris$Petal.Length,  
xlab="Sepal length (in cm)", ylab="Petal length (in cm)",  
main="Comparison between sepal and petal length of 50 iris", pch=20)
```

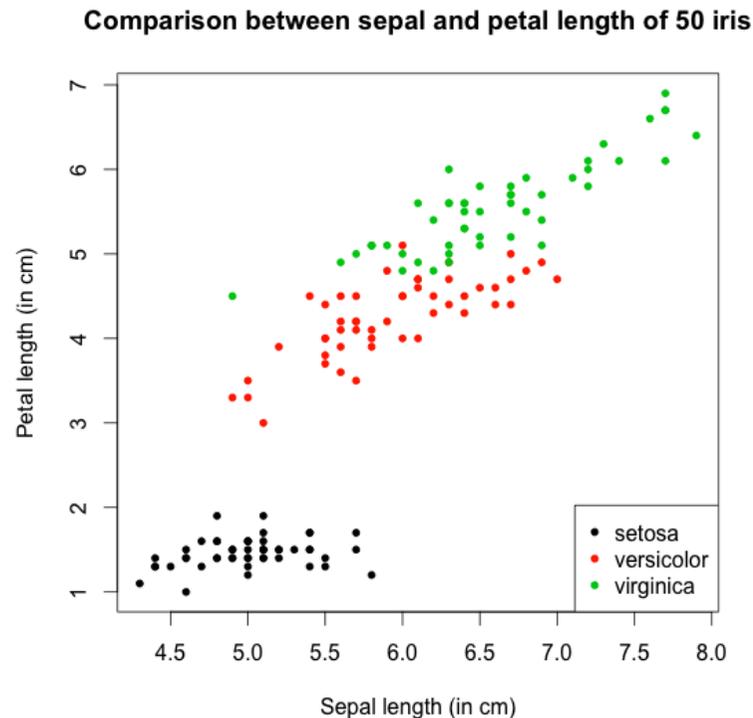


Exercise 5 :

scatterplot with different colours

```
# graph
plot(iris$Sepal.Length, iris$Petal.Length,
     xlab="Sepal length (in cm)", ylab="Petal length (in cm)",
     main="Comparison between sepal and petal length of 50 iris", pch=20,
     col=iris$Species)

# legend
legend("bottomright", legend=levels(iris$Species), col=1:3, pch=20)
```



Exercise 6

```
# Mean number of protein coding genes per chromosome
```

```
mean(huge$Protein.coding.genes )
```

```
[1] 879.5
```

```
# Maximum number of protein coding genes per chromosome
```

```
max(huge$Protein.coding.genes)
```

```
[1] 2153
```

```
# The above information can also be obtained using the summary function
```

```
summary(huge$Protein.coding.genes)
```

```
Min. 1st Qu.  Median   Mean 3rd Qu.  Max.
```

```
80.0  626.2  884.0  879.5 1106.0 2153.0
```

```
# Chromosome with the highest number of coding genes
```

```
huge[huge$Protein.coding.genes ==max(huge$Protein.coding.genes ),1]
```

```
[1] 1
```

```
# Chromosome with the smallest number of coding genes
```

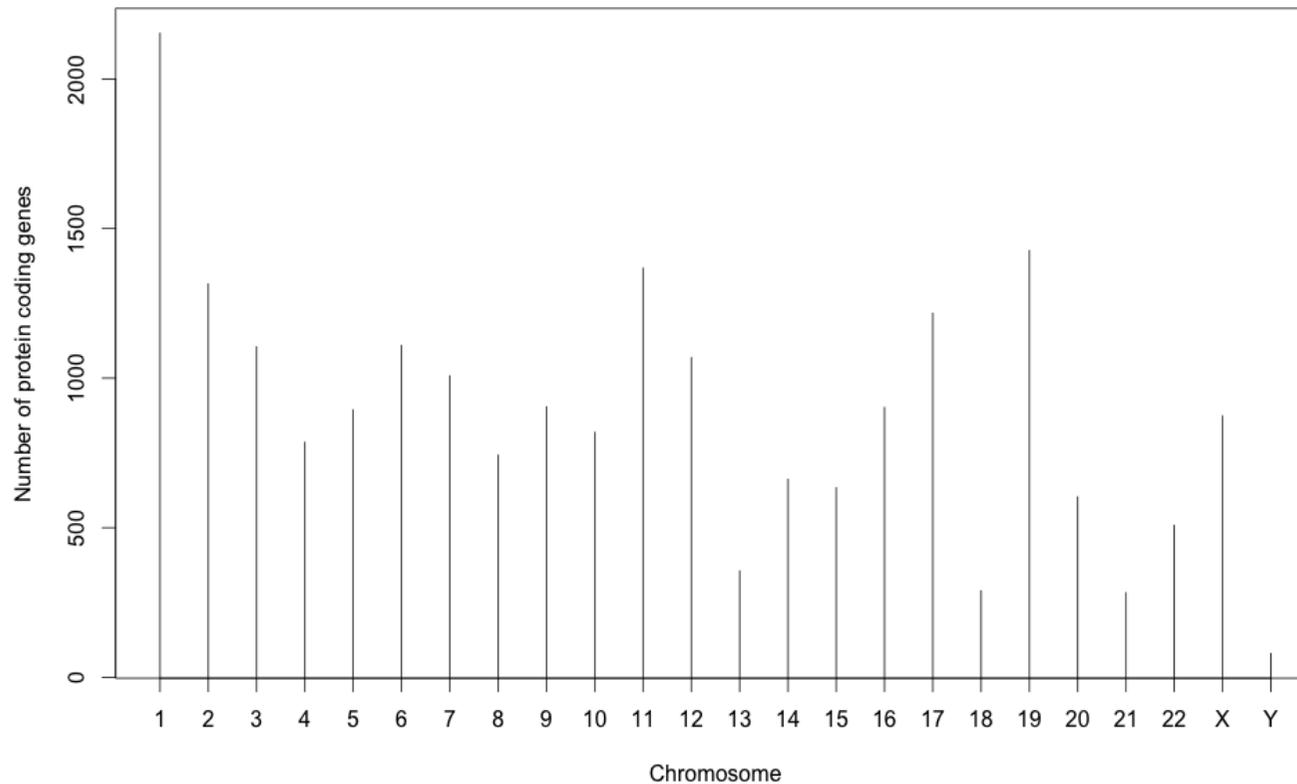
```
huge[huge$Protein.coding.genes ==min(huge$Protein.coding.genes ),1]
```

```
[1] Y
```

Exercise 6

Bar-chart

```
plot(huge$Protein.coding.genes, xlab="Chromosome",  
ylab="Number of protein coding genes", type="h", xaxt="n")  
axis(1, at=1:length(huge$Chr), labels=huge$Chr)
```



Exercise 7

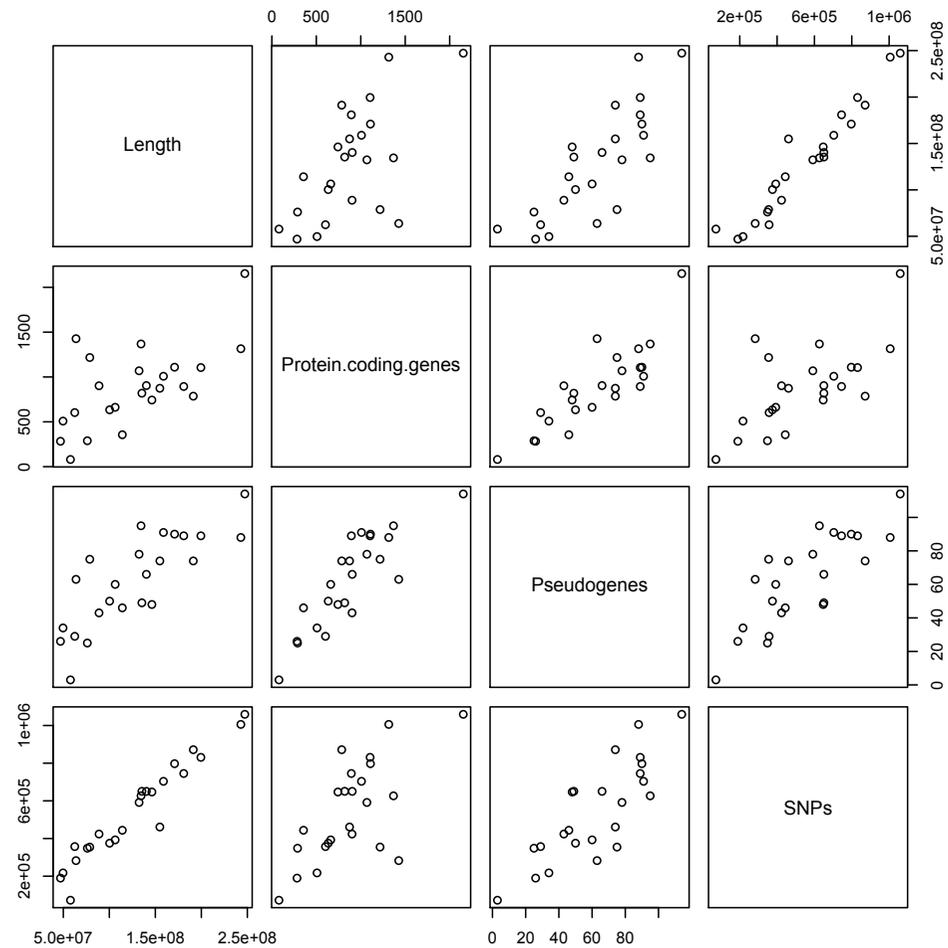
```
# Pearson correlation coefficient  
# between chromosome size and the number of protein coding genes  
cor(huge$Length, huge$Protein.coding.genes)  
[1] 0.6305566
```

```
# Pearson correlation coefficients between all pairs of quantitative variables  
cor(huge[,2:5])
```

	Length	Protein.coding.genes	Pseudogenes	SNPs
Length	1.0000000	0.6305566	0.8070697	0.9631934
Protein.coding.genes	0.6305566	1.0000000	0.8527515	0.6731252
Pseudogenes	0.8070697	0.8527515	1.0000000	0.8158501
SNPs	0.9631934	0.6731252	0.8158501	1.0000000

Exercise 7

Graphical representation
pairs(huge[,2:5])

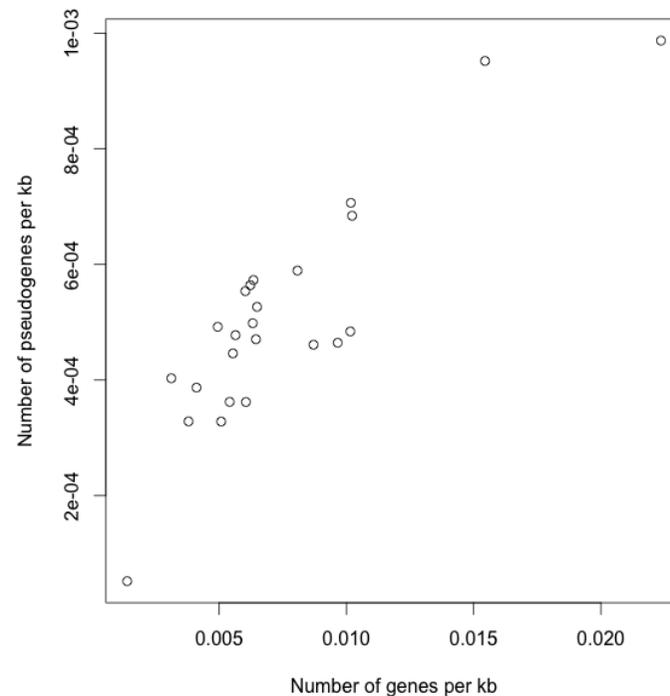


Exercise 7

```
# Calculation of the number of genes, pseudogenes and SNP per kb for each chromosome  
hugeNorm = huge[,3:5]/huge[,2]*1000
```

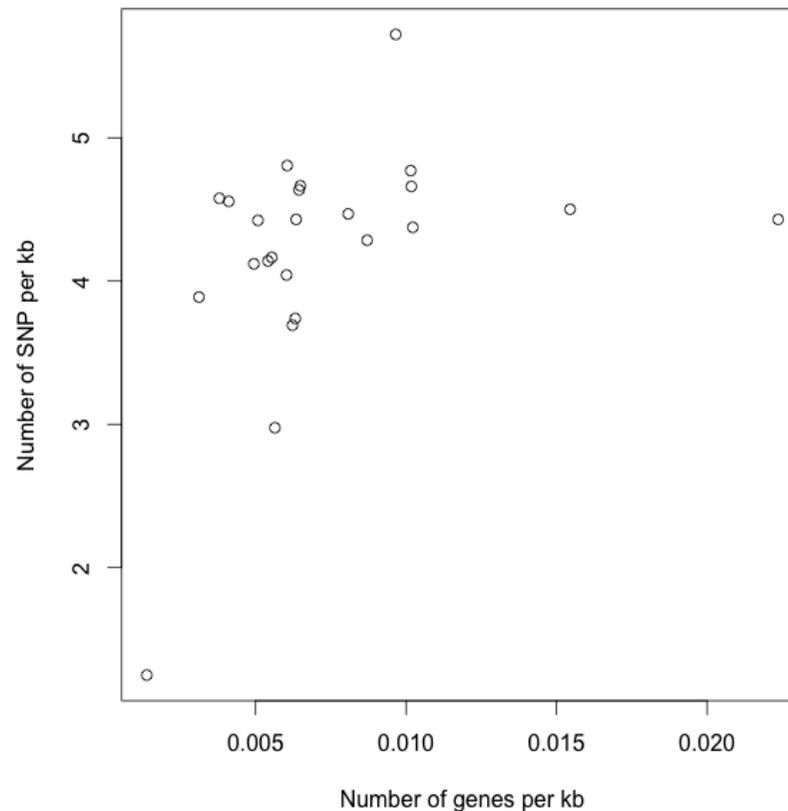
```
# Correlation between genes and pseudogenes numbers per kb  
cor(hugeNorm$Protein.coding.genes, hugeNorm$Pseudogenes)  
[1] 0.87249
```

```
plot(hugeNorm$Protein.coding.genes, hugeNorm$Pseudogenes,  
xlab="Number of genes per kb",ylab="Number of pseudogenes per kb")
```



Exercise 7

```
# Correlation between SNP and genes numbers  
cor(hugeNorm$Protein.coding.genes, hugeNorm$SNPs)  
[1] 0.3849074  
plot(hugeNorm$Protein.coding.genes, hugeNorm$SNPs,  
xlab= "Number of genes per kb",ylab= "Number of SNP per kb")
```



Exercise 8

```
molecule = read.table("molecule.txt", header=TRUE, sep="\t")
```

```
# Wilcoxon test
```

```
# H0 : the blood level of this molecule does not increase significantly after the treatment
```

```
# H1 : the blood level of this molecule increases significantly after the treatment
```

```
# Let  $\alpha=0.05$ 
```

```
wilcox.test(molecule$After, molecule$Before, alternative="greater", paired=TRUE)
```

```
Wilcoxon signed rank test
```

```
data: molecule$After and molecule$Before
```

```
V = 20, p-value = 0.03125
```

```
alternative hypothesis: true location shift is greater than 0
```

```
# Conclusion :
```

```
We reject H0, the blood level of this molecule significantly increases after the treatment
```