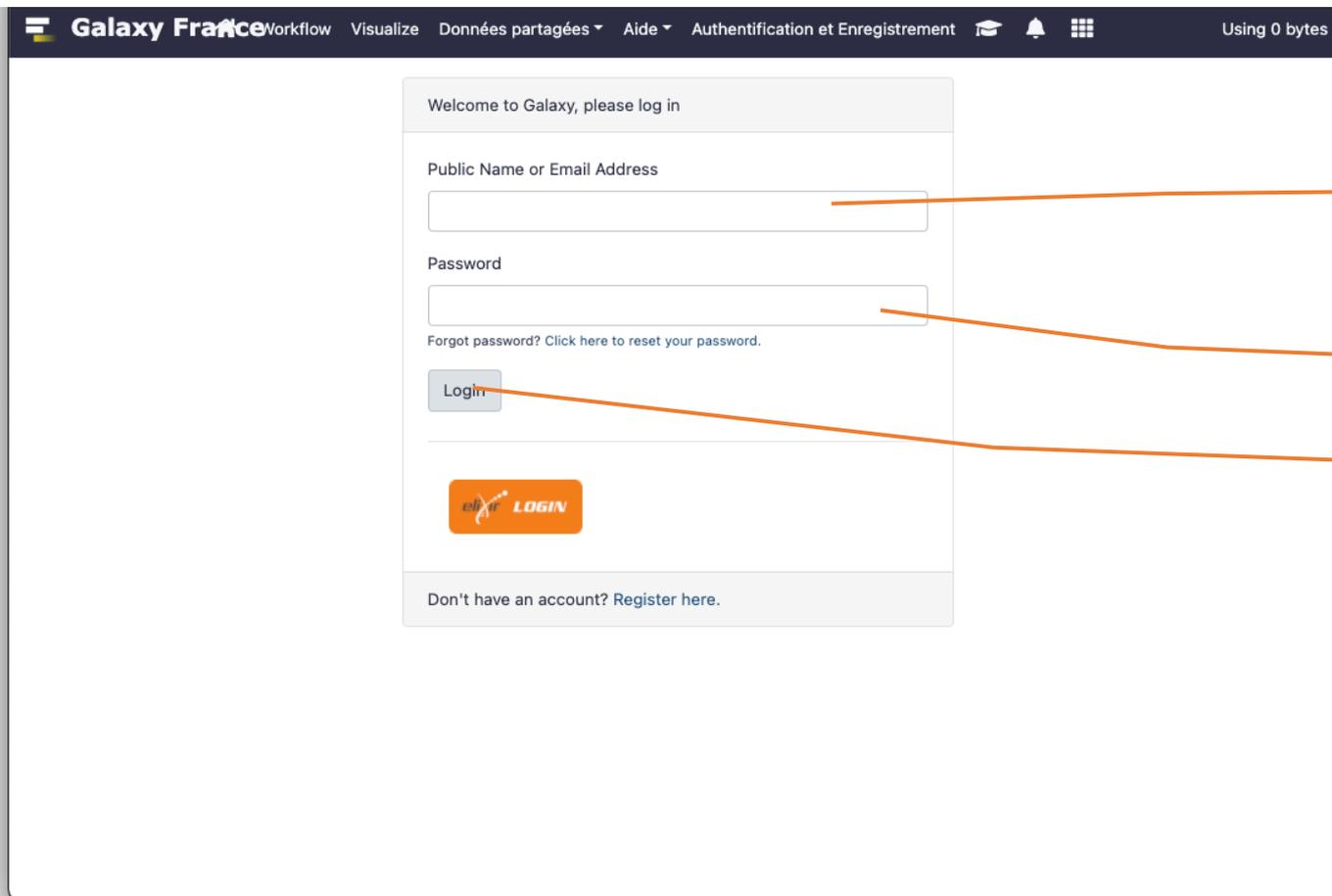


Introduction to Galaxy (answers to questions)

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Answer 1 : Log in

- Go to <https://usegalaxy.fr/>



The screenshot shows the Galaxy France login interface. At the top, there is a dark navigation bar with the text "Galaxy France" and several menu items: "Workflow", "Visualize", "Données partagées", "Aide", and "Authentification et Enregistrement". On the right side of the navigation bar, it says "Using 0 bytes". Below the navigation bar, the main content area is white and contains a login form. The form has a header "Welcome to Galaxy, please log in". It includes two input fields: "Public Name or Email Address" and "Password". Below the password field, there is a link: "Forgot password? Click here to reset your password." There is a "Login" button and a larger orange "LOGIN" button with a logo. At the bottom of the form, there is a link: "Don't have an account? Register here." Three orange lines point from the right side of the image to the input fields and the "Login" button.

Enter your login

Enter your password

Click on Login

Answer 2 : History

- Create a new history



- Change the name of the new history to “RNA-seq data analysis” by clicking on “Unnamed history” on top of the history panel. Then type “RNA-seq data analysis” and [ENTER].



Answer 3 : Import data to Galaxy

- 1.
 - Go to Shared data (top menu) > Histories.
 - Search « NGS data analysis training Strasbourg ».

The screenshot shows the Galaxy France interface. The top navigation bar includes 'Galaxy France', 'Workflow', 'Visualize', 'Données partagées', 'Aide', 'Authentification et Enregistrement', and 'Using 0%'. The left sidebar contains a 'Tools' section with a search bar and an 'Upload Data' button. Below this is a list of tool categories, including 'Get Data', 'Collection Operations', 'GENERAL TEXT TOOLS', 'Text Manipulation', 'Filter and Sort', 'Join, Subtract and Group', 'GENOMIC FILE MANIPULATION', 'Convert Formats', 'FASTA/FASTQ', 'FASTQ Quality Control', 'SAM/BAM', 'BED', 'VCF/BCF', 'Nanopore', 'COMMON GENOMICS TOOLS', 'Operate on Genomic Intervals', 'Fetch Alignments/Sequences', and 'GENOMICS ANALYSIS'. An orange arrow points to the search bar in the 'Tools' section, which contains the text 'NGS data analysis training Strasbourg'. The main content area displays a table of 'Published Histories' with columns for 'Name', 'Annotation', and 'Owner'. The first entry is 'NGS data analysis training Strasbourg' by 'stephanie'. The right sidebar shows an empty 'History' section with a message: 'Cet historique est vide. You can Charger vos propres données or Charger des données depuis une source externe'.

Name	Annotation	Owner
NGS data analysis training Strasbourg		stephanie
galaxy101		lgueuen
Rustenholtz - GW		dc244f268e
RNASeq GW BV - Rustenholtz		dc244f268e
RNASeq RI MIM - Rustenholtz		dc244f268e
RNASeq RI BV - Rustenholtz		dc244f268e
Rustenholtz - RI		dc244f268e
ABIMS - Galaxy initiation 2022 - datasets - public		lgueuen
Rustenholtz - IGV		dc244f268e
Rustenholtz - DE Sangiovese		dc244f268e
Rustenholtz - DE Barbera Refosco		dc244f268e
RNASeq GW MIM - Rustenholtz		dc244f268e
GCMS_For_WF		yguilton44
TP RNAseq billie Initial datasets		c0d111ceec2
ARTIC v3	Auxiliary datasets for analysis of ARTIC (v3) SARS-CoV-2 data	lecorguille

Answer 3 : Import data to Galaxy

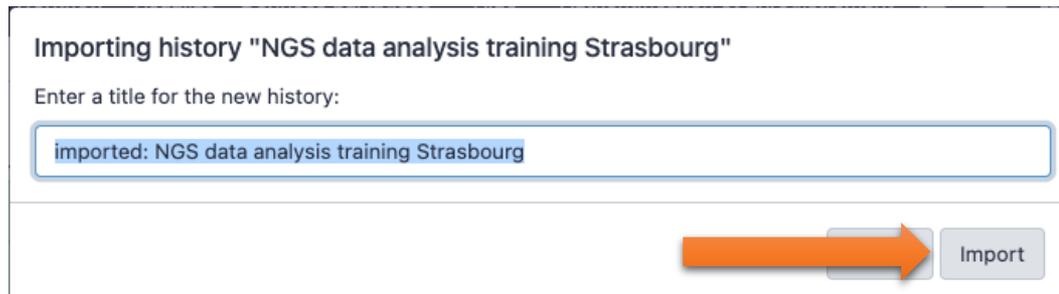
- Click on « **NGS data analysis training Strasbourg** »
- Click on 



The screenshot shows the Galaxy France interface. At the top, there is a navigation bar with the logo and menu items: Workflow, Visualize, Données partagées, Aide, and Authentification et Enregistrement. The main content area displays a history titled "NGS data analysis training Strasbourg" with a size of 2 GB. Below the title is a search bar labeled "Rechercher des données". Underneath, there are two data files listed in a table:

Jeu de données	Annotation
2: siLuc3_S12040.fastq.gz	
1: siLuc2_1000000.fastq.gz	

On the right side, there is a sidebar with a plus icon at the top, an author profile for "stephanie" with a blue geometric logo, and a "Related Histories" section. An orange arrow points to the plus icon in the top right corner of the history card.



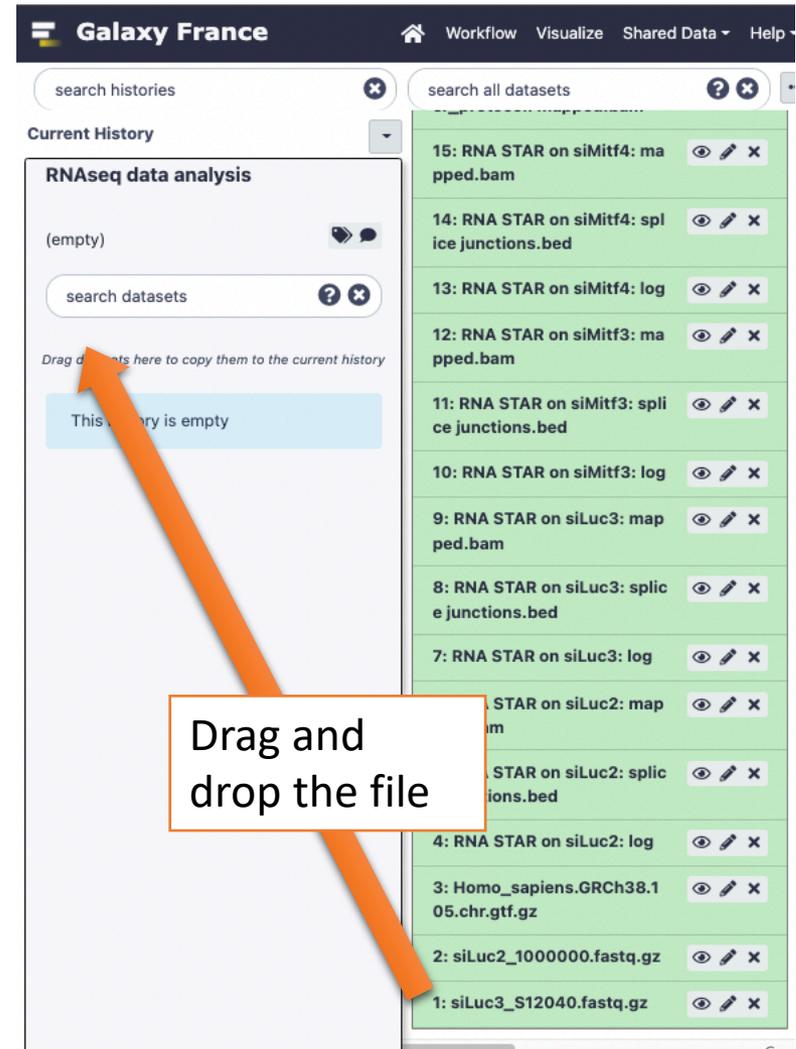
The screenshot shows a dialog box titled "Importing history 'NGS data analysis training Strasbourg'". It prompts the user to "Enter a title for the new history:" and has a text input field containing "imported: NGS data analysis training Strasbourg". At the bottom right, there is an "Import" button. An orange arrow points to the "Import" button.

- Click on Import

Answer 3 : Import data to Galaxy

- Use the tool "View all histories" 

then:



The screenshot displays the Galaxy France web interface. At the top, there is a navigation bar with the text "Galaxy France" and several icons. Below the navigation bar, there are two search bars: "search histories" and "search all datasets". The main content area is divided into two panels. The left panel, titled "Current History", shows an empty history with the text "(empty)" and a "search datasets" search bar. Below this, there is a blue box with the text "This history is empty". The right panel shows a list of datasets, numbered 1 to 15, with their names and file extensions. An orange arrow points from a text box to the "search datasets" search bar in the left panel.

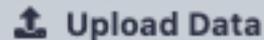
Drag and drop the file

Dataset ID	Dataset Name	File Extension
15	RNA STAR on siMitf4: mapped	.bam
14	RNA STAR on siMitf4: splice junctions	.bed
13	RNA STAR on siMitf4: log	
12	RNA STAR on siMitf3: mapped	.bam
11	RNA STAR on siMitf3: splice junctions	.bed
10	RNA STAR on siMitf3: log	
9	RNA STAR on siLuc3: mapped	.bam
8	RNA STAR on siLuc3: splice junctions	.bed
7	RNA STAR on siLuc3: log	
6	RNA STAR on siLuc2: mapped	.bam
5	RNA STAR on siLuc2: splice junctions	.bed
4	RNA STAR on siLuc2: log	
3	Homo_sapiens.GRCh38.105.chr.gtf	.gz
2	siLuc2_1000000.fastq	.gz
1	siLuc3_S12040.fastq	.gz

Answer 3 : Import data to Galaxy

• 2

- Click on the button to display the drag and drop utility

 Upload Data

- Drag and drop the file [sample.bed.gz](#) into the opened window.
 - **Type:** bed
 - **Genome:** Mouse July 2007 (NCBI37/mm9) (mm9)

Download from web or upload from disk

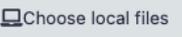
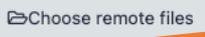
Download from web or upload from disk

Regular Composite Collection Rule-based

You added 1 file(s) to the queue. Add more files or click 'Start' to proceed.

Name	Size	Type	Genome	Settings	Status
 sample.bed.gz	482.9 KB	bed	Mouse July 200...		0% 

Type (set all): Auto-detect Genome (set all): unspecified (?)

   **Start** Pause Reset Close

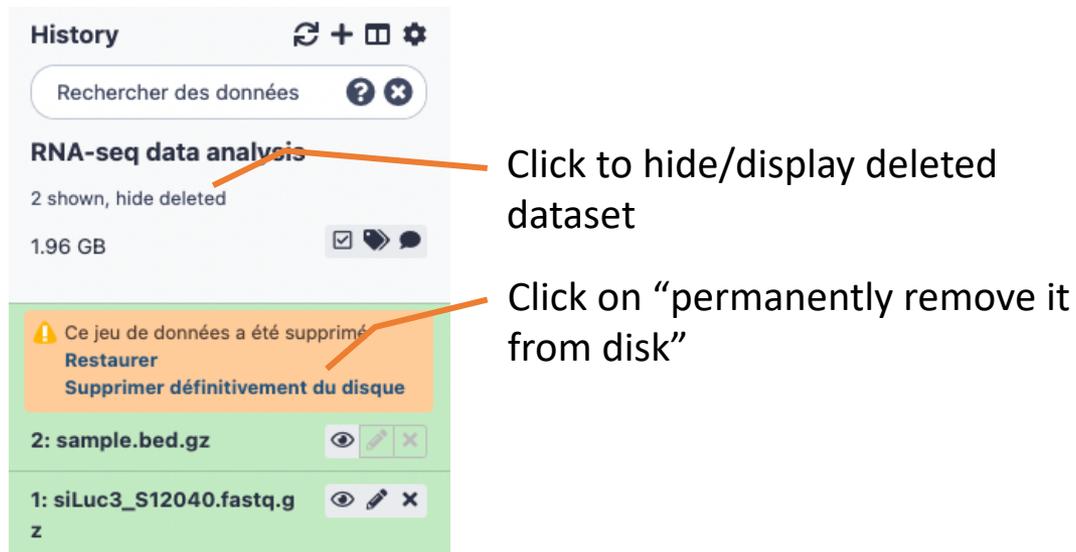
- Click on Start
- Click on Close to close the upload utility

Answer 4: remove dataset

- 1. Click on the  of the dataset in the history



- 2.
 - A)



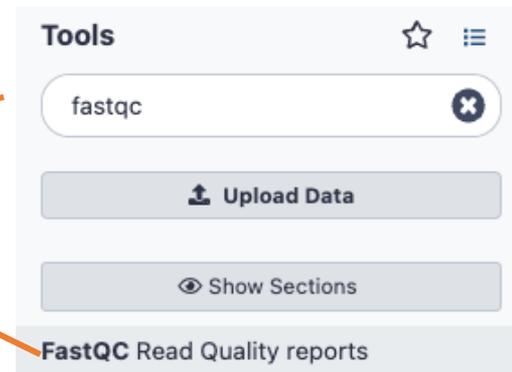
- B) Click on "hide deleted"
- NOTE: to permanently remove from disk multiple deleted datasets, click on the History option button  and select "Purge deleted datasets"

Answer 5 : Running a tool

1. Search “fastqc” in the list or using the search field of the tool panel.

Enter: “fastqc”

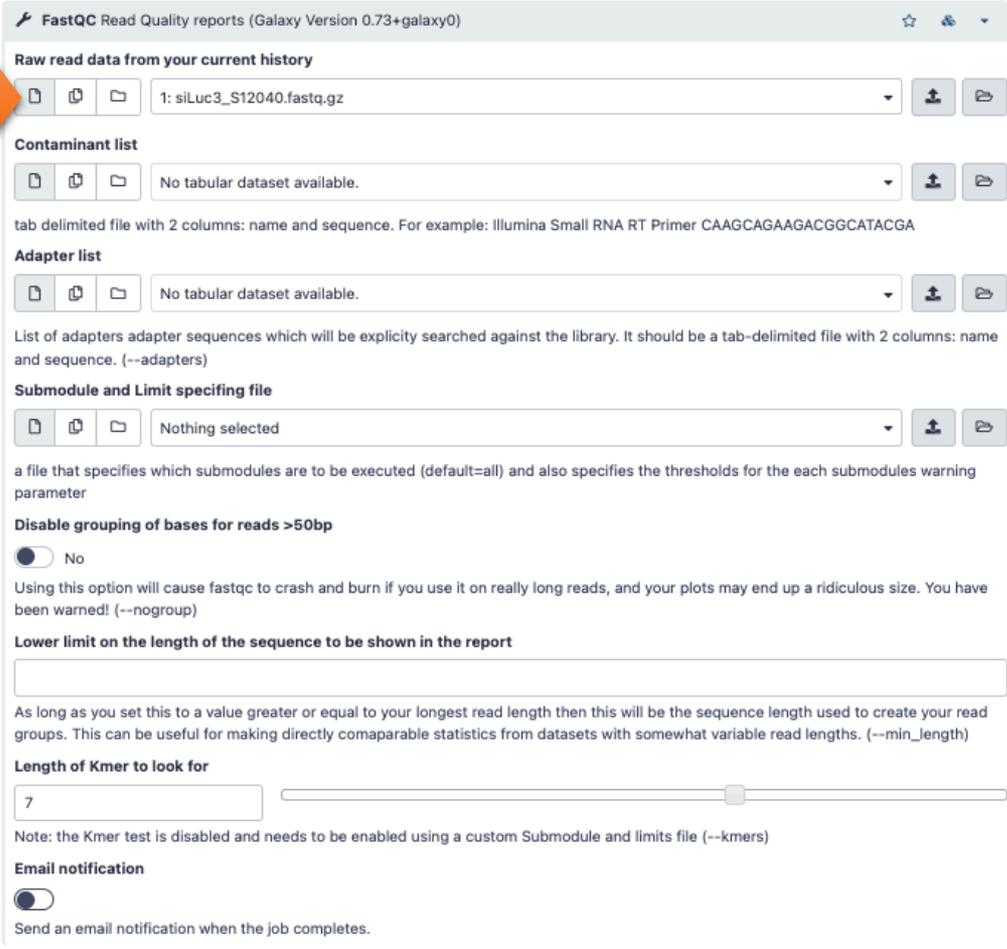
Click on the tool name



Answer 5 : Running a tool

2. Select the file to analyze and click on Execute

1. 



FastQC Read Quality reports (Galaxy Version 0.73+galaxy0)

Raw read data from your current history

1: siLuc3_S12040.fastq.gz

Contaminant list

No tabular dataset available.

tab delimited file with 2 columns: name and sequence. For example: Illumina Small RNA RT Primer CAAGCAGAAGACGGCATACGA

Adapter list

No tabular dataset available.

List of adapters adapter sequences which will be explicitly searched against the library. It should be a tab-delimited file with 2 columns: name and sequence. (--adapters)

Submodule and Limit specifying file

Nothing selected

a file that specifies which submodules are to be executed (default=all) and also specifies the thresholds for the each submodules warning parameter

Disable grouping of bases for reads >50bp

No

Using this option will cause fastqc to crash and burn if you use it on really long reads, and your plots may end up a ridiculous size. You have been warned! (--nogroup)

Lower limit on the length of the sequence to be shown in the report

As long as you set this to a value greater or equal to your longest read length then this will be the sequence length used to create your read groups. This can be useful for making directly comparable statistics from datasets with somewhat variable read lengths. (--min_length)

Length of Kmer to look for

7

Note: the Kmer test is disabled and needs to be enabled using a custom Submodule and limits file (--kmers)

Email notification

Off

Send an email notification when the job completes.

2. 