Introduction to R software

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Introduction to R software

- R software fundamentals
- Graphics using R
- Basic statistics using R

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- Graphics using R
- Basic statistics using R



- Created by Ross Ihaka and Robert Gentleman*
 - https://www.r-project.org/
- R is
 - An environment allowing to perform
 - statistical analyses
 - graphics
 - A programming language
 - Inspired by S
- Freely distributed under GNU General Public License

Where to download R?

- The comprehensive R archive network (CRAN)
 - http://cran.r-project.org
 - Available for Linux, Mac and Windows

The Comprehensive R Archive Network

Download and Install R
Precompiled binary distributions of the base system and contributed packages,
Windows and Mac users most likely want one of these versions of R:

Download R for Linux
Download R for (Mac) OS X
Download R for Windows

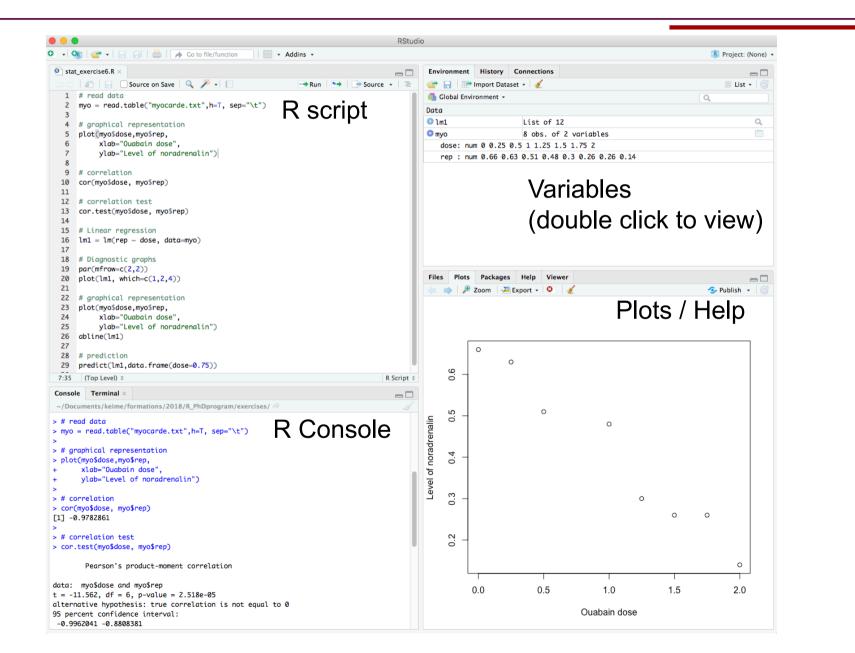
R is part of many Linux distributions, you should check with your Linux package

- RStudio
 - https://www.rstudio.com/
 - Integrated development environment for R
 - Available for Linux, Mac and Windows (https://www.rstudio.com/products/rstudio/download/)

management system in addition to the link above.



RStudio



Start an R session

- Launch R or RStudio
- Choose a working directory
 - Why ?
 - Store data files : by default R will search files in this directory
 - Save results : graphs, files, R working sessions
 - Continue a previous analysis
 - How ?
 - > setwd("path/to/directory")
 - Rstudio : Session / Set Working Directory / Choose Directory
 - Drag and drop your folder on RStudio icon
 - Verification
 - > getwd() : get current working directory
 - RStudio : Console title

→ Create a working directory for this training and choose to work in this directory

→ Verify that you are working in the correct directory

Interactive usage of R (1/2)

• Principle

- Enter a command and press return
- R execute the command (and eventually display the corresponding result)
- Another command can then been entered
- Examples
 - > 1+1

[1] 2

Comment (not taken into account by R)

- > x=1 # affectation
- > <u>x</u> [1] 1
- [1] 1

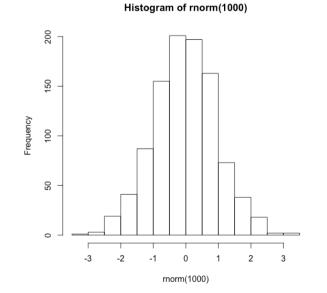
The arrow **↑** allow to recall a previous command

Interactive usage of R (2/2)

• Examples

> rnorm(10) # rnorm(): function, c.f. brackets
[1] -1.087784299 1.157426928 0.364218560
[4] 2.065149368 -0.893725503 -0.251256812
[7] 0.005916529 0.072525550 -0.446474935
[10] -0.399408315

> hist(rnorm(1000)) #rnorm() : graphical function



Backups

- Backups during an interactive session

 \rightarrow Prefer to save commands in an R script (text file with R commands AND comments)

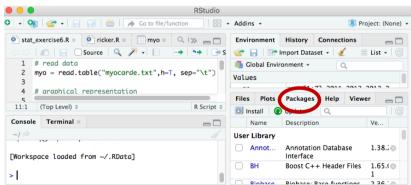
• Save when exiting R

>q() # to exit R (or click on Save workspace image? [y/n/c]: y

- Continue an analysis started with R
 - > load("path/to/.RData")
 - or open R in the directory chosen for this analysis (e.g. drag and drop this directory to RStudio icon)

R libraries (1/2)

- A lot of functions available by default in R (base library)
- More specific functions (multivariate analyses, clustering, non linear regression, phylogeny, …) → libraries
 - A library is a set of tools on a specific subject
 - RStudio : packages tab
 - Several libraries are installed within R :
 - > installed.packages() # list of all installed packages and their version
 - > library() #list of installed libraries and brief description
 - To install a new library
 - > install.packages("library_name")
 - Or click on install on RStudio *Packages* tab
 - To load a library in the working session
 - > library(library_name)
 - Required to be able to use a function from the package *library_name*, in each R session



R libraries (2/2)

- CRAN libraries :
 - https://cran.r-project.org/ Packages link
 - 12,308 libraries currently available
- Bioconductor project :
 - http://www.bioconductor.org/
 - Provides tools for the analysis of high-throughput genomic data
 - Last version : 3.6
 - 1,473 packages to analyse and represent genomic data
 - 911 annotations packages
 - 326 packages containing experiment data
 - Installation
 - source("http://bioconductor.org/biocLite.R")
 - biocLite() #install core packages
 - biocLite("library_name") #install a specific package called library_name

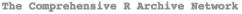


Task Views Search About R R Homepage The R Journal Software R Sources R Binaries

Packages

What's new?

CRAN Mirrors



Download and Install R

Precompiled binary distributions of the base system and contributed packages, **Windows and Mac** users most likely want one of these versions of R:

- Download R for Linux
- Download R for (Mac) OS X

Download R for Windows

R is part of many Linux distributions, you should check with your Linux package management system in addition to the link above.

Source Code for all Platforms

Windows and Mac users most likely want to download the precompiled binaries listed in the upper box, not the source code. The sources have to be compiled before you can use them. If you do not know what this means, you probably do not want to do it!

• The latest release (2018-03-15, Someone to Lean On) R-3.4.4.tar.gz,



Sources of documentation on R (1/2)

- R homepage : http://www.r-project.org
 - Manuals (to start : An introduction to R) + contributed documentation link
 - FAQ
- Rseek : http://www.rseek.org
- R Site Search : http://finzi.psych.upenn.edu/search.html
- Bioconductor : http://bioconductor.org/help/
 - Very helpful : package vignettes and workflows
- Within R :
 - help.start() # html help / Help tab on RStudio
 - ?y # help on y
 - Very useful when you know the name of the object/function of interest
 - Example : ?plot
 - ??subject
 - # help pages containing the word "subject"# Search Engine & Keywords in RStudio
 - Helpful when you don't know the name of the corresponding object/function
 - Example : ??Student

Files Plots Packages Help Image: Image and the second seco	Viewer C						
Statistical Data Analysis							
Manuals							
An Introduction to R Writing R Extensions R Data Import/Export							
Reference							
Packages	Search Engine & Keywords						
Miscellaneous Material							
About R License NEWS	Authors Resources Frequently Asked Questions Thanks User Manuals Technical papers						

Sources of documentation on R (2/2)

- Structure of an R help page (c.f. for example ?plot)
 - Тор
 - Name of the help page and corresponding library
 - Description
 - Brief description of the utility of the function/operator
 - Usage
 - Syntax to use in order to call a function (with default parameters values)
 - Typical use of an operator
 - Arguments
 - For each argument of the function : detailed description and type
 - Details
 - Precisions on the function/operator (possibly references)
 - Value
 - If applicable, the type of the object returned by the function
 - See also
 - Other help pages related to this page
 - Examples
 - Examples of use of the function
 - Can be executed with the example() function : c.f. > example(plot)

Objects

- Variables, data, results are stored in memory in objects
- We act on these objects using **operators** and **functions**
- Object name
 - Must start with a letter and may contain : a-z / A-Z / 0-9 / . / _
 - Case sensitive (a \neq A)
- An object can be created with the affectation operator := or <- > a = 1
- To view the content of an object : type its name
 a

 [1] 1
- List all objects in memory : ls()
 > ls()
 [1] "a"
- Remove an object from memory : rm()

> rm(a) # remove from memory the object a
> ls()
character(0)

Operators

- Main arithmetic operators
 - + * / ^(power)
- Relational operators
 - < > <= >= == (equality) != (difference)
- Logical operators
 - No : !
 - And : &
 - Or : |

Functions

- Call a function : function name()
- Function arguments
 - Into brackets
 - − Argument not given → default value if defined, c.f. ?function
 - Arguments identification
 - By name. Can be abbreviated if the abbreviation is not ambiguous
 - By position

Example :

```
> ?seq
> seq(from=1, to=10, by=1)
[1] 1 2 3 4 5 6 7 8 9 10
> seq(fr=1, to=10, by=1)
[1] 1 2 3 4 5 6 7 8 9 10
> seq(1, 10, 1)
[1] 1 2 3 4 5 6 7 8 9 10
> seq(to=10, by=1)
[1] 1 2 3 4 5 6 7 8 9 10
# identification by position
# identification by position
# from=1 by default
```

Some useful functions in R

- To obtain information about an object
 - class : class of the object
 - length resp. dim : dimension of an object with 1 resp. 2 dimensions
 - head : the beginning of an object (useful for the visualisation of large objects)
 - summary : summary of the content of an object
- Mathematical functions
 - log, log10, log2, exp
 - min, max : minimal and maximal values of an object / range : c(min,max)
 - sum, diff, prod : sum, difference and product of the elements of a vector
 - round : round of a value
- Objects manipulation
 - sort : sort a vector or factor
 - order : give the permutation which allows to sort a vector

```
> obs = c(3,9,7,4,1)
> obs
[1] 3 9 7 4 1
> sort(obs)
[1] 1 3 4 7 9
> order(obs)
[1] 5 1 4 3 2
```

Some useful functions in R : examples

iris is a dataset available in R

It contains the measurements (in cm) of sepal length and width and petal length and width, for 50 flowers from 3 species of iris (*Iris setosa, versicolor* and *virginica*).

> data(iris) # load the iris dataset

> class(iris) #class of the iris object

[1] "data.frame"

> dim(iris) # dimensions of the dataset (number of lines and columns)

[1] 150 5

> head(iris) # visualize the beginning of this dataset

Sepal.Length Sepal.Width Petal.Length Petal.Width Species

1	5.1	3.5	1.4	0.2 setosa
2	4.9	3.0	1.4	0.2 setosa
3	4.7	3.2	1.3	0.2 setosa
4	4.6	3.1	1.5	0.2 setosa
5	5.0	3.6	1.4	0.2 setosa
6	5.4	3.9	1.7	0.4 setosa





from https://fr.wikipedia.org/wiki/Iris_de_Fisher

Exercise 1

Use the *iris dataset* available in R :

- → load iris dataset with > load(iris)
- → How many flowers from setosa species are available in this dataset ?
- → Print sepal length of all flowers in increasing order
- → What is the mean sepal length of all flowers ?
- → How many flowers have a sepal length larger than 5 cm ?
- → How many *setosa* flowers have a sepal length larger than 5 cm ?

Note :

- > iris\$Sepal.Length # column Sepal.Length from the iris dataset
- > sum(Y) where Y is a logical vector gives the number of TRUE elements in this vector

Class of an object

- Different object classes depending on what and how we want to store values
 - Vectors : numeric, character, logical
 - Factors : factor
 - Arrays : data.frame or matrix
 - Lists : list
 - Functions : function
- To know the class of an object use the class() function

Vectors

- A vector contains one or several values of the same type
 - numeric : numbers
 - character : character strings
 - logical : Boolean values (TRUE or FALSE)
 - → Even the smallest objects (e.g. only 1 element) are vectors in R

• Creation of a vector

```
With several numbers, characters or boolean values : c()
obs = c(3,9,7,4,1)
obs
[1] 3 9 7 4 1
Sequence of integers with an increment of 1 or -1 : :
v = 1:5
v
[1] 1 2 3 4 5
```

Numeric vectors content

- Integers
- Real numbers
 - With a decimal point : 0.2
 - Scientific notation : 2e-1 # corresponds to 2 x 10⁻¹
- Infinite : Inf et –Inf
 > 1/0
 > -1/0
 [1] Inf
 [1] –Inf
- Not a number : NaN
 - > 0/0 [1] NaN > log(-1) [1] NaN
- Missing values : NA

Logical vectors

- A logical vector may contain 3 different values
 - TRUE or T
 - FALSE or F
 - NA (missing value)
- Often created from an operation

> obs = c(3,9,7,4,1)
> obs>3 #observations strictly higher than 3
[1] FALSE TRUE TRUE TRUE FALSE

Character strings vectors

- A character strings vector may contain
 - One or more strings delimited by " "
- Several useful functions to manipulate character strings
 - paste
 - To concatenate several strings
 - grep
 - Search for an expression in character strings
 - substr
 - Extract or replace a substring in a vector of character strings
 - match (or %in%)
 - x %in% y return TRUE for each element of x present in y
 - unique(x)
 - Return one time each distinct element from x

Indexation of a vector

•	Operator [] allows to access elements from a vector	> v = 1:5
	> v[2]	> v
	[1] 2	[1] 1 2 3 4 5
	> v[6] # Return NA if we exceed the length of the object	

• Several possible indexations

[1] NA

- With a vector of **positive integers** : indices of the elements to keep
 v[2:4]

 [1] 2 3 4
- With a vector of negative integers : indices of the elements to exclude
 > v[-c(1,5)]
 [1] 2 3 4
- → It is impossible to specify both positive and negative indexes
- With a logical vector : TRUE correspond to the values to keep, FALSE to the values to exclude

```
> v[ c(FALSE, TRUE, TRUE, TRUE, FALSE) ]
[1] 2 3 4
```

Very useful to keep only the elements from a vector satisfying a given criterion :
> iris\$Sepal.Length[iris\$Species=="setosa"] #Sepal Length from iris setosa

Manipulation of vectors

- To know the number of elements in a vector : length()
 > length(v)
 [1] 5
- Operators and usual mathematical functions operate element by element Examples :

```
> v
[1] 1 2 3 4 5
> v2 = 5:1
> v2
[1] 5 4 3 2 1
> v + v2
[1] 6 6 6 6 6
> v+1
[1] 2 3 4 5 6
> 2 * v
[1] 2 4 6 8 10
> v==5
[1] FALSE FALSE FALSE FALSE TRUE
```

Exercise 2

Use the *iris* dataset available in R

- > sl = iris\$Sepal.Length
- What is the class of sl object ?
- What is the minimal and maximal sepal length of these flowers ?
- What is the length of the 10 largest sepals ?

Factors

- Factors allow to store qualitative variables in R
- A factor in R is made of :
 - The values of a qualitative variable
 - The levels of this variable, i.e. the different possible values of this variable, eventually ordered
- Use factor() function to create factors in R

Creation of factors

Non-ordered qualitative variables

```
e.g. the winter behaviour of bird species : migratory (M), partial migratory (PM) or sedentary (S)
```

```
> wb = factor(c("M","M","S", "PM","S")) # winter behaviour of 5 species
> wb
[1] M M S MP S
Levels: M MP S
> class(wb)
[1] "factor"
```

- Ordered qualitative variables
 - specify : ordered = TRUE in the factor creation instruction
 - If the argument levels are not specified, the alphabetical order is used

e.g. the diversity of bird species observed in several stations : low, medium or high

```
> diversity = factor(c("low","high","medium","medium","low"), levels = c("low","medium","high"),
ordered = T) # diversity of bird species in 5 stations.
```

```
# the alphabetical order is not appropriate → we need to precise the levels > names(diversity) = paste("s",1:5,sep="") # names of the corresponding stations
```

> diversity

s1s2s3s4s5lowhighmediummediumlowLevels:low < medium < high</td>

Manipulation of factors

- Levels of a factor : levels()
 - > levels(wb)
 - [1] "M" "PM" "S"
 - > levels(diversity)
 - [1] "low" "medium" "high"
- Contingency table (counts for each factor level) : table()
 - table(wb)
 wb
 M PM S
 2 1 2
 table(diversity)
 diversity
 low medium high
 2 2 1
- Extract element(s) from a factor : []
 - Same principle as for a vector

Exercise 3

Use the *iris* dataset available in R

- What is the class of the "Species" object ?
- What are the different iris species in this dataset?
- How many flowers from each species are present in this dataset ?
- What is the species of the iris with the smallest sepal length ?

Lists (1/2)

- Very flexible data structure :
 - List = several elements of any type
 - An element from a list can be any object : vector, factor, list, data.frame, matrix, function
- Creation of a list : list()

mylist = list(elt1=value1, elt2=value2, ...)

- elt1, elt2 : name of each element in the list (not required)
- value1 : value of the first element (any R object)
- Extract element(s) from a list :
 - mylist\$elt1 only if a name has been set to the element in the list
 - elt1 : name of the element in the list that we want to extract
 - Use names(mylist) to know the names of the elements from a list
 - mylist[[i]]
 - To extract the element i from the list mylist

Lists (2/2)

R functions often return lists

➔ It allows to easily search for information among all the results provided by R Example :

```
> s1 = rnorm(100)
> s2 = rnorm(100)
```

```
# Student's t-test
```

```
> ttest = t.test(s1,s2,var.equal=T)
```

> ttest

Two Sample t-test data: ech1 and ech2 t = 0.8984, df = 198, p-value = 0.3701 alternative hypothesis: true difference in means is not equal to 0 95 percent confidence interval: -0.1445087 0.3863550 sample estimates: mean of x mean of y 0.116775078 -0.004148088 # t.test function return many information # ttest is a list
> is.list(ttest)
[1] TRUE

names of the elements in the list ttest
> names(ttest)
[1] "statistic" "parameter" "p.value"
[4] "conf.int" "estimate" "null.value"
[7] "alternative" "method" "data.name"

extract the p-value of the test
> ttest\$p.value
[1] 0.3700663

extract the confident interval
> ttest\$conf.int
[1] -0.1445087 0.3863550
attr(,"conf.level")
[1] 0.95

Matrices

- Table (2 dimensions) that contain elements of the same type
 → We can create a matrix of mode numeric, character or logical
 > m = matrix(1:6, nrow=2, ncol=3)
 > m

 [,1] [,2] [,3]
 [1,] 1 3 5
 [2,] 2 4 6
 > dim(m) # number of rows and columns
 [1] 2 3
- To extract elements from a matrix : [
 - To extract the value in line i and column j : matrix[i,j] > m[2,1]
 [1] 2
 - To extract all rows : matrix[,j] > m[,3]
 [1] 5
 - To extract all columns : matrix[i,] > m[2,]

6

- Mathematical operators and functions work element by element
 - > m-m [,1] [,2] [,3] [1,] 0 0 0 [2,] 0 0 0

data.frame

- data.frame : a table
 - Made of several numeric, logical and/or character vectors
 - All these vectors must have the same length
 - Difference from matrix : vectors could be of different types
- Often the most appropriate class to store datasets
 - > class(iris)
 [1] "data.frame"
 > dim(iris) # dimensions of *iris* data.frame
 [1] 150 5
 # *iris* data.frame contains vectors of different types :
 > class(iris\$Sepal.Length)
 [1] "numeric"
 > class(iris\$Species)
 - [1] "factor"
- Usually data are available in a file
 - Use the read.table function to create a data.frame by reading data from this file

Manipulation of data.frame

• Indexation

- As for matrices
 - df[i,j] to extract the value in line i and column j
 - Example :

> head(iris)

Sepal.Length Sepal.Width Petal.Length Petal.Width Species

1	5.1	3.5	1.4	0.2 setoso	1				
2	4.9	3.0	1.4	0.2 setoso	1				
3	4.7	3.2	1.3	0.2 setoso	1				
4	4.6	3.1	1.5	0.2 setoso	1				
5	5.0	3.6	1.4	0.2 setoso	1				
6	5.4	3.9	1.7	0.4 setoso	1				
> iris[3,2]									
[1] 3.2	2								

- You can also use the operator \$ to extract a column
 - > iris\$Sepal.Length #same as iris[,1]
- Dimensions
 - dim() function

Data input and output

- File format for data input : tabulated text
 - Data in excel
 - Save as text
 - Choose a field separator that is never used in your data, e.g. tabulation
- Most useful function for data input : read.table()
 - Read a text file and create a data.frame with the data from this file
 - Rows and columns from the created data.frame correspond to rows and columns from the file

Data input

- Main options of the read.table() function (c.f. ?read.table)
 - file
 - Name of the file (between quotes, or a character string object)
 - By default R search for this file in the working directory (otherwise specify the corresponding path)
 - header
 - TRUE or FALSE depending on whether the first row contains header information
 - sep
 - Field separator (e.g. "\t" for tabulations)
 - quote
 - · Characters used to delimit the character variables
 - dec
 - Character used for decimal points ("." by default)
 - row.names
 - Vector with the names of rows
 - Or number of the column that contains row names in the file (e.g. 1)

Data input example

- The ecrin.txt file is available on the following webpage : http://genomeast.igbmc.fr/wiki/doku.php?id=training:introduction2r
- This file contains the number of bird species (RIC column) observed in Ecrins national park in 1991 (1315 observations)*
- Each observation was done
 - In a station indicated in the STA column
 - During the week indicated in the SEM column
 - Either during the morning or the afternoon
 - HEU column : 1:morning, 2:afternoon

Data input example

read ercrin.txt file and put the corresponding data.frame in the ecrin object # in the ecrin.txt file the first line correspond to the column names > ecrin=read.table("ecrin.txt", header=TRUE)

```
> class(ecrin)
[1] "data.frame"
```

```
> head(ecrin)
  STA SEM HEU RIC
     2 1
  3
            5
 1
  3 2 2 3
 2
 3 3 3 1 5
  3 3 2 3
4
  3 4 1
 5
            4
 6
         2
  3
      4
            1
```

You can then manipulate the corresponding data using R
e.g. select only observations performed during the morning
> ecrin.morning = ecrin[ecrin\$HEU == 1,]

Save data into a file (1/2)

- write.table() function
 - Write an object (vector, data.frame or matrix) in a file
- Main options of the function write.table()
 - 1st argument
 - · Name of the object to write
 - file
 - Name of the file in which you want to write (default output to the R console)
 - append
 - If TRUE append to the file
 - default=FALSE : any existing file with this name is destroyed
 - quote
 - If TRUE (default), character and factor columns, row and column names will be surrounded by " "
 - sep
 - The field separator string

Save data into a file (2/2)

- Main options of the function write.table()
 - na
 - The string to use for missing data
 - dec
 - The string to use for decimal points
 - row.names / col.names
 - whether the row/column names of the data.frame are to be written (TRUE or FALSE)
 - or a character vector of row/column names to be written

• Example

- # Creation of a file ecrin_morning.txt containing only observations performed during the morning > write.table(ecrin.morning, file="ecrin_morning.txt", quote=F, sep="\t", row.names=FALSE)
- # options :
- Do not use quote for character and factor columns
- Use tabulation as the field separator string
- Do not write line numbers

Exercise 4

For this exercise use the dataset available in the humanGenomeSummary.txt file. This file contains for each human chromosome its length in bp, the number of coding genes, pseudogenes and SNP.

This file is available on the following webpage :

http://genomeast.igbmc.fr/wiki/doku.php?id=training:introduction2r

- Import this file into R
- What is the total number of protein coding genes?
- Retrieve the chromosomes with more than 1,000 annotated protein coding genes annotated.
- Create a file containing chromosome number and number of protein coding genes, ordered by decreasing protein coding gene number.

Conditional execution : make choices

- Allow to execute R instructions only under certain conditions
- Syntax

 if (condition){
 true.block
 }
 else{
 false.block
 }
 - → If condition is true, true.block is executed, otherwise false.block is executed

Loops : repeat an action (1/2)

- Allow to iteratively perform several analyses
- For loop

```
    Syntax
    for (variable in suite) {
    expression
    }
```

→ Execute *expression* iteratively for each value of *variable* contained in *suite*

• While loop

```
    Syntax
    while (condition){
    expression
    }
```

→ Execute *expression* while *condition* is true

Introduction to R software

- R software fundamentals
- Graphics using R
- Basic statistics using R

Introduction to graphics with R

- Data visualisation
 - An essential step in the exploratory data analysis
- R
 - A flexible and powerful way to perform graphics
 e.g. demo(graphics) or http://rgraphgallery.blogspot.fr
- Principles
 - Graphics functions
 - To perform graphics
 - Options of graphics functions and graphics parameters
 - To modify graphics presentation
 - Graphics devices
 - In which all graphics operations occur : window or file

Dataset for example plots

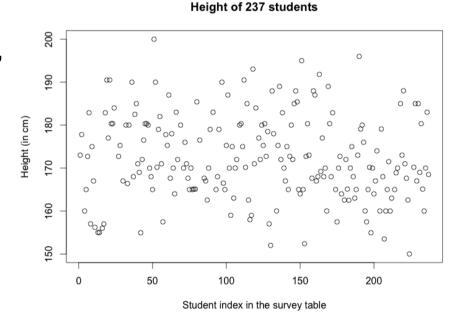
library(MASS) data(survey) ?survey #contains the responses of 237 students #at the University of Adelaide to a number of questions head(survey)

	Sex	Wr.Hnd	NW.Hnd	W.Hnd		Fol	d	Pulse	Clap	Exer	Smoke	Height	M.I	Age
1	Female	18.5	18.0	Right	R	on	L	92	Left	Some	Never	173.00	Metric	18.250
2	Male	19.5	20.5	Left	R	on	L	104	Left	None	Regul	177.80	Imperial	17.583
3	Male	18.0	13.3	Right	L	on	R	87	Neither	None	0ccas	NA	<na></na>	16.917
4	Male	18.8	18.9	Right	R	on	L	NA	Neither	None	Never	160.00	Metric	20.333
5	Male	20.0	20.0	Right	Nei	the	r	35	Right	Some	Never	165.00	Metric	23.667
6	Female	18.0	17.7	Right	L	on	R	64	Right	Some	Never	172.72	Imperial	21.000

Quantitative variable in R : numeric vector

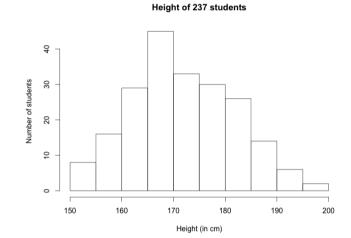
Scatterplot : plot()
 plot(survey\$Height,
 main="Height of 237 students",
 xlab="Student index in the survey table",
 ylab="Height (in cm)"
)

... not very informative...



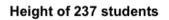
Histogram : hist()

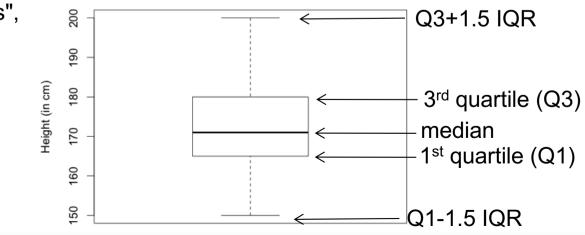
 hist(survey\$Height,
 main="Height of 237 students",
 xlab="Height (in cm)",
 ylab="Number of students")



– Boxplot : boxplot()

boxplot(survey\$Height, main="Height of 237 students", ylab="Height (in cm)")



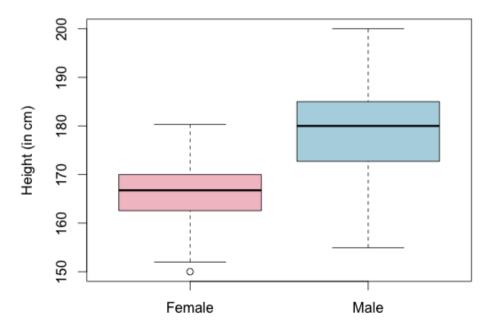


IQR=interquartile range (Q3-Q1) \rightarrow contains 50% of the individuals

- Boxplots allow to easily compare distributions

```
boxplot(survey$Height ~ survey$Sex,
    main="Height of 237 students",
    ylab="Height (in cm)",
    col=c("pink","lightblue")
```

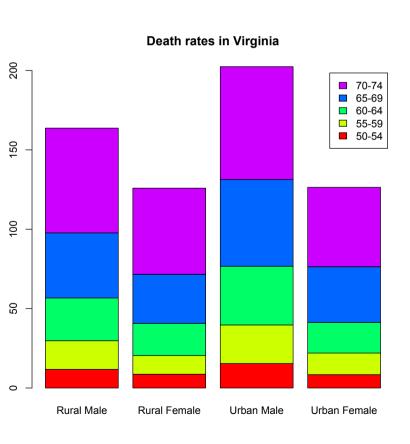
Height of 237 students



- barplot()

VADeaths # Death rates per 1000 in Virginia in 1940									
	Rural	Male	Rural	Female	Urban	Male	Urban	Female	
50-54		11.7		8.7		15.4		8.4	
55-59		18.1		11.7		24.3		13.6	
60-64		26.9		20.3		37.0		19.3	
65-69		41.0		30.9		54.6		35.1	
70-74		66.0		54.3		71.1		50.0	

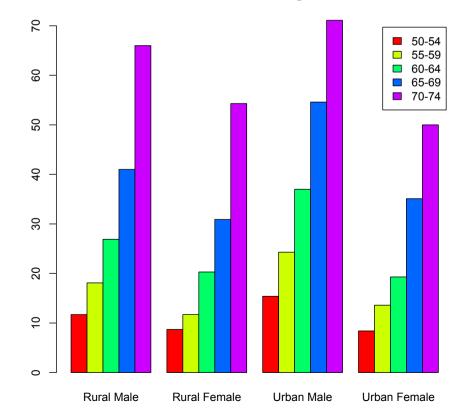
barplot(VADeaths, legend=rownames(VADeaths), col=rainbow(5), main="Death rates in Virginia")



- barplot()

barplot(VADeaths, legend=rownames(VADeaths), col=rainbow(5), main="Death rates in Virginia", beside=TRUE)

Death rates in Virginia



– barplot2() : in order to represent confidence intervals

myci.l = VADeaths*0.9 # just for the example...

myci.u = VADeaths*1.1

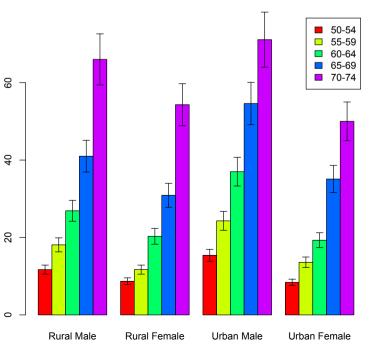
library(gplots)

barplot2(VADeaths, legend=rownames(VADeaths),

col=rainbow(5), main="Death rates in Virginia", beside=TRUE,

ci.l=myci.l, ci.u=myci.u, plot.ci=TRUE)

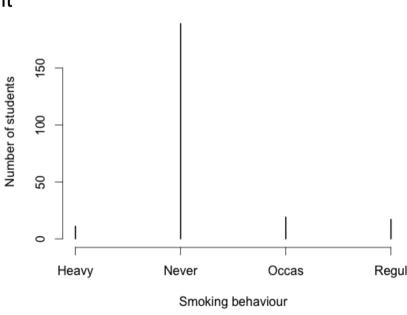
Death rates in Virginia



- Bar-chart (vertical lines) : plot(...,type="h")

plot(table(survey\$Smoke), type="h", main="Smoking behaviour of 237 students", ylab= "Number of students", xlab= "Smoking behaviour")

N.B. : type="h" is the default type to represent contingency tables (plot.table)

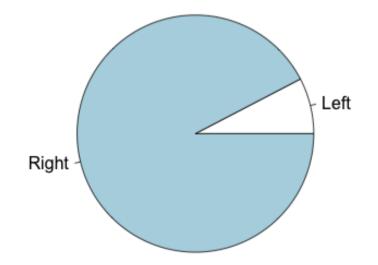


Smoking behaviour of 237 students

• Pie-chart

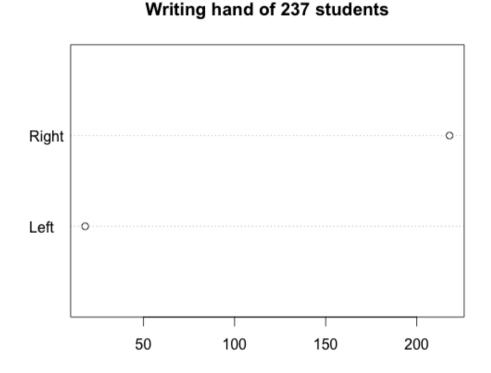
pie(table(survey\$W.Hnd), main= "Writing hand of 237 students")

Writing hand of 237 students

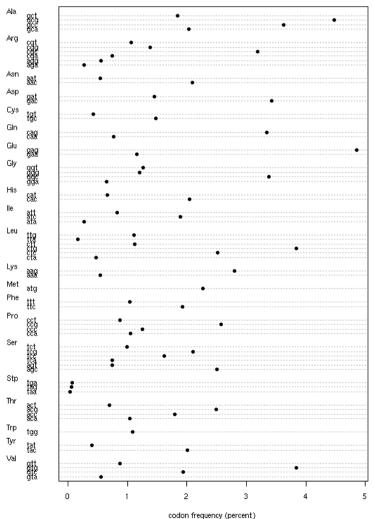


• Cleveland's dot plot

dotchart(table(survey\$W.Hnd), main= "Writing hand of 237 students")



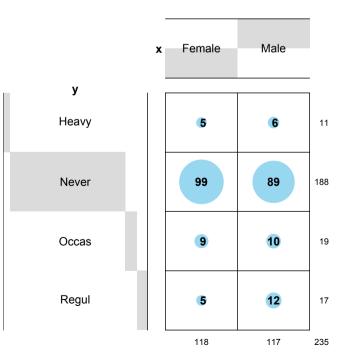
Very useful for variables with a lot of levels, e.g. codon usage :



- Qualitative qualitative
 - contingency table : balloonplot()

library(gplots)
balloonplot(table(survey[,c("Sex","Smoke")]))

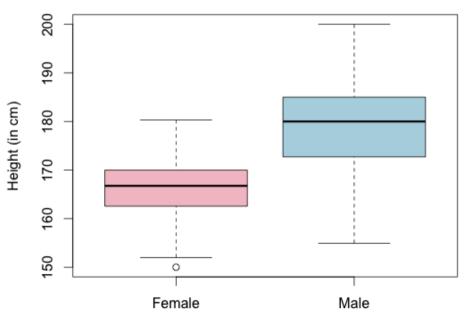
Balloon Plot for x by y. Area is proportional to Freq.



- Quantitative qualitative
 - boxplot()

```
boxplot(survey$Height ~ survey$Sex,
    main="Height of 237 students",
    ylab="Height (in cm)",
    col=c("pink","lightblue")
```

Height of 237 students

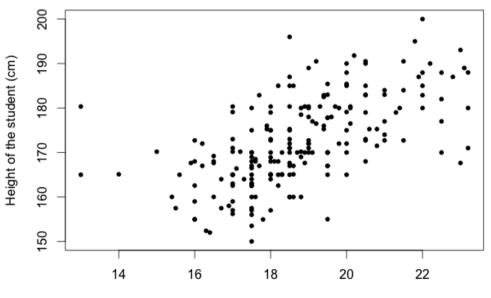


• Quantitative – quantitative

- scatter plot : plot()

plot(survey\$Wr.Hnd, survey\$Height, pch=20, xlab= "Span of writing hand (cm)", ylab= "Height of the student (cm)", main= "Height as a function of span of writing hand for 237 students")

Height as a function of span of writing hand for 237 students

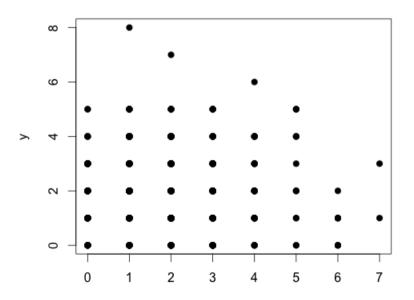


• Quantitative – quantitative

```
- Overlapping points problem
```

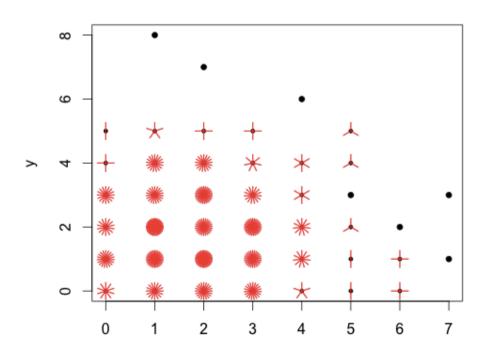
```
n = 500
x = rpois(n, lambda = 2)
y = rpois(n, lambda = 2)
plot(x, y, pch = 19, main = "A misleading scatter plot")
```

A misleading scatter plot



- Quantitative quantitative
 - Overlapping points problem

sunflowerplot(x, y, pch = 19, main = "With sunflowerplot()")

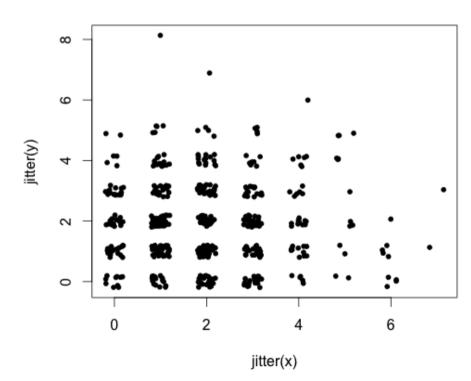


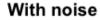
With sunflowerplot()

Multiple points are plotted as 'sunflowers' with multiple leaves

- Quantitative quantitative
 - Overlapping points problem

plot(jitter(x), jitter(y), main = "With noise", pch=20)





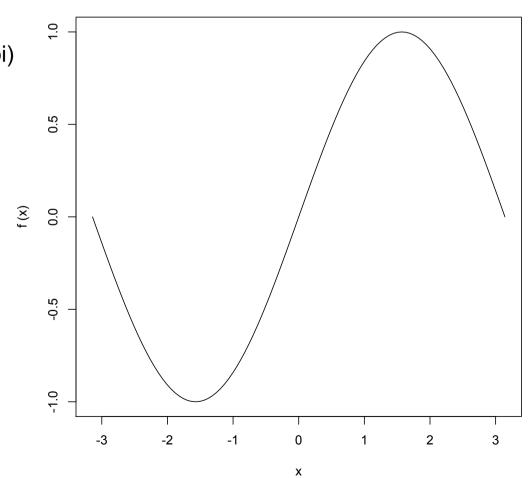
A small amount of noise is added to the data

Curves

• Draw functions

- curve()

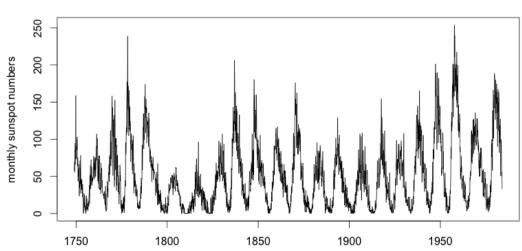
f = function(x){ sin(x) }
curve(f, from = -pi, to = pi)



Curves

- Evolution of a variable as a function of a second variable
 - plot(..., type="l")

e.g. time course data data(sunspots) plot(time(sunspots), sunspots, type="l", main = "Monthly sunspot numbers from 1749 to 1983", xlab= "Year", ylab="monthly sunspot numbers")



Monthly sunspot numbers from 1749 to 1983

Functions to modify plots

- Add points to a plot : points()
- Add lines to a plot
 - lines()
 - abline()
 - a,b : to specify intercept and slope
 - h : to specify y-value(s) for horizontal line(s)
 - v : to specify x-value(s) for vertical line(s)
- Add a legend : legend()
- Add a title : title()
- Add axes : axis()
- Add text to a plot : text(x, y, labels)
 - x,y : the coordinates where text labels should be written
- Draw segment(s) or arrow(s) : segments(), arrows()

Functions to modify plots

Example : add a legend to a graph

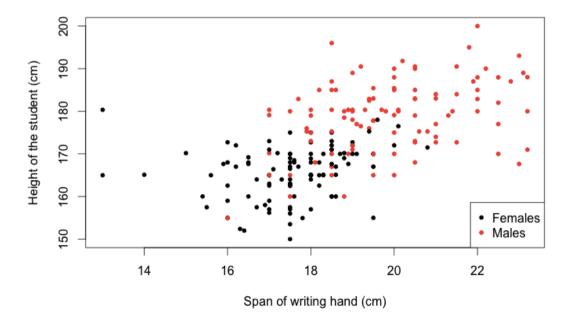
plot(survey\$Wr.Hnd,survey\$Height, pch=20,

xlab= "Span of writing hand (cm)",

ylab= "Height of the student (cm)",

main= "Height as a function of span of writing hand for 237 students",col=survey\$Sex)

legend("bottomright", legend=c("Females", "Males"), col=c(1,2), pch=20)



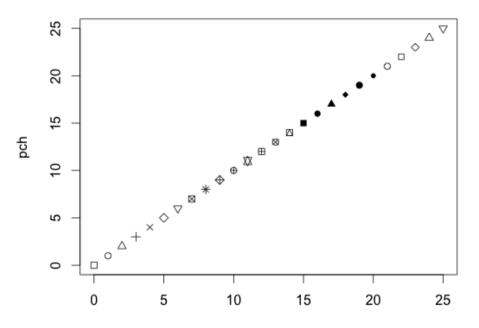
Height as a function of span of writing hand for 237 students

- c.f. help page of each graphics function
 - A lot of different arguments
- Common arguments for several graphics functions
 - main : title
 - sub : subtitle
 - xlab, ylab : axes annotation
 - xlim, ylim : minimal and maximal values for the axes
 - type : "p" : points, "l" : lines, "b" : points+lines, "n" : no plotting,
 "h" : vertical lines
 - pch : plotting character(s) or symbol(s)
 - Ity : line type(s), lwd : line width(s)
 - col : color(s), colors() list all available colours

- Different plotting characters and symbols
 - 0:25 : R symbols

plot(0:25,0:25,pch=0:25, xlab="pch",ylab="pch",main="Different R symbols")

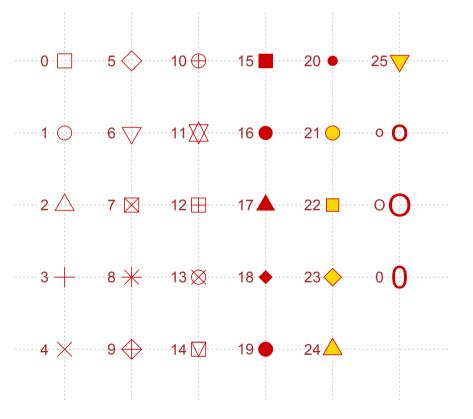
- 32:127 : ASCII characters



Different R symbols

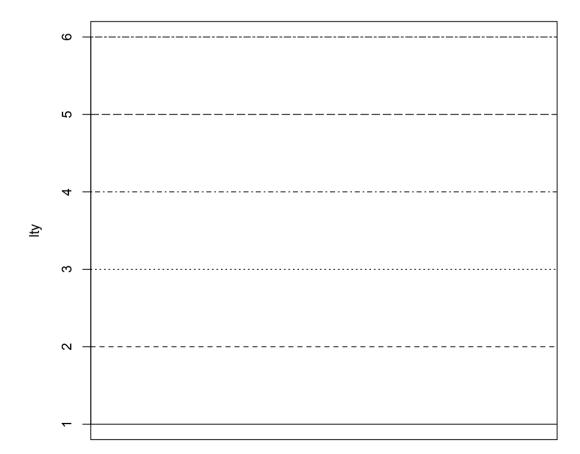
?points

```
##----- Showing all the extra & some char graphics symbols ------
pchShow <-
 function(extras = c("*",".", "o","0","0","+","-","|","%","#"),
           cex = 3, ## good for both .Device=="postscript" and "xll"
           col = "red3", bg = "gold", coltext = "brown", cextext = 1.2,
           main = paste("plot symbols : points (... pch = *, cex =",
                        cex,")"))
  {
    nex <- length(extras)</pre>
    np <- 26 + nex
    ipch <- 0:(np-1)
   k <- floor(sqrt(np))</pre>
    dd < -c(-1,1)/2
    rx \le dd + range(ix \le ipch %/% k)
    ry <- dd + range(iy <- 3 + (k-1)- ipch %% k)
    pch <- as.list(ipch) # list with integers & strings
    if(nex > 0) pch[26+ 1:nex] <- as.list(extras)
    plot(rx, ry, type="n", axes = FALSE, xlab = "", ylab = "",
         main = main)
    abline(v = ix, h = iy, col = "lightgray", lty = "dotted")
    for(i in 1:np) {
      pc <- pch[[i]]
      ## 'col' symbols with a 'bg'-colored interior (where available) :
      points(ix[i], iy[i], pch = pc, col = col, bg = bg, cex = cex)
      if(cextext > 0)
         text(ix[i] - 0.3, iy[i], pc, col = coltext, cex = cextext)
    }
 }
pchShow()
```

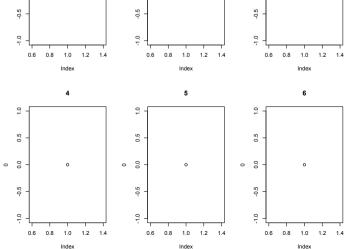


• Different lines

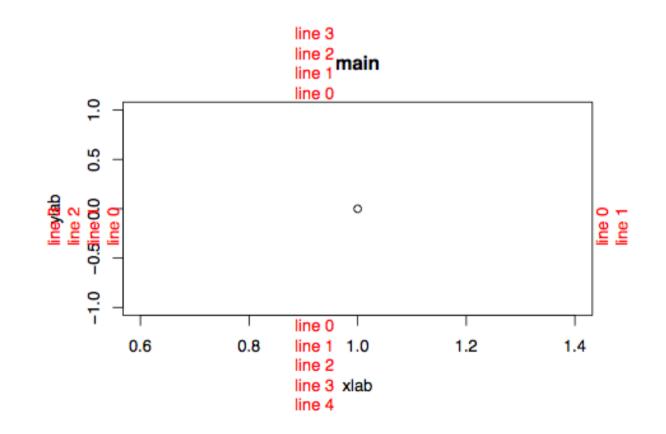
plot(1:6,1:6,type="n",xaxt="n",ylab="lty",xlab="")
for (i in 1:6){abline(h=i,lty=i)}



- All graphical parameters
 - ?par # how to set or query graphical parameters
 - par() # list of the 71 graphical parameters and their values
- Main graphical parameters
 - Several graphics in the same window : mfrow = c(nrows, ncolumns) par(mfrow = c(2, 3)) for (i in 1:6) plot(0, main = i)

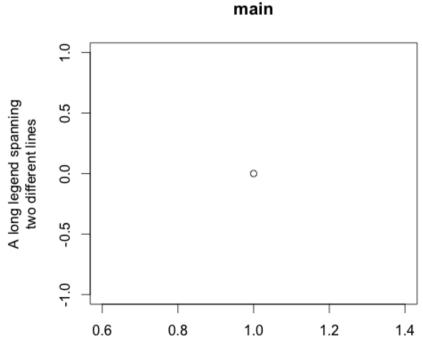


- Margins : mar = c(bottom, left, top, right)
 - In text lines
 - By default, mar = c(5,4,4,2) + 0.1



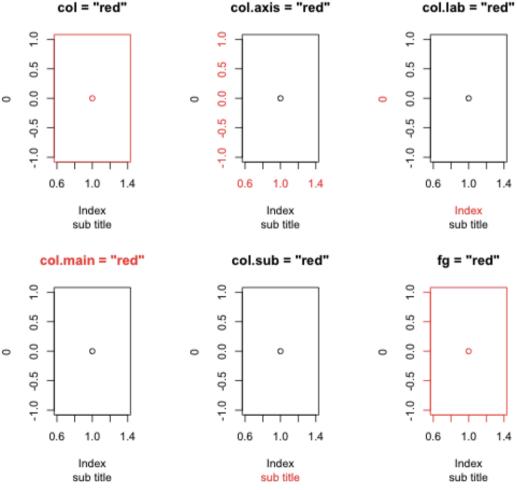
– Margins : mar = c(bottom, left, top, right)

Example : add a margin line to the left margin : par(mar = par("mar") + c(0, 1, 0, 0)) plot(0, xlab = "xlab", ylab = "A long legend spanning\ntwo different lines", main = "main")



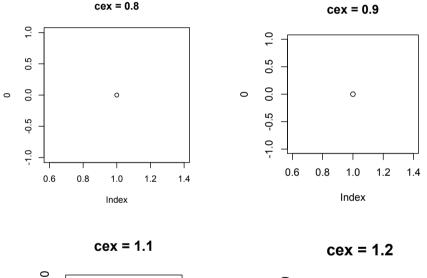
– Colours of the elements in a graphics : col*

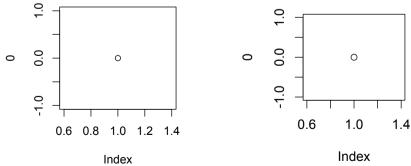
```
par(mfrow = c(2, 3))
par(col = "red")
plot(0, main = "col = \"red\"", sub = "sub title")
par(col = "black", col.axis = "red")
plot(0, main = "col.axis = \"red\"", sub = "sub title")
par(col.axis = "black", col.lab = "red")
plot(0, main = "col.lab = \"red\"", sub = "sub title")
par(col.lab = "black", col.main = "red")
plot(0, main = "col.main = \"red\"", sub = "sub title")
par(col.main = "col.sub = \"red\"", sub = "sub title")
plot(0, main = "col.sub = \"red\"", sub = "sub title")
plot(0, main = "col.sub = \"red\"", sub = "sub title")
plot(0, main = "fg = \"red\"", sub = "sub title")
```



```
- Relative size of characters : cex*
par(mfrow = c(2, 2))
for (cex in seq(from = 0.8, to = 1.2, length = 4)) {
par(cex = cex)
plot(0, main = paste("cex =", round(cex, 1)))
}
```

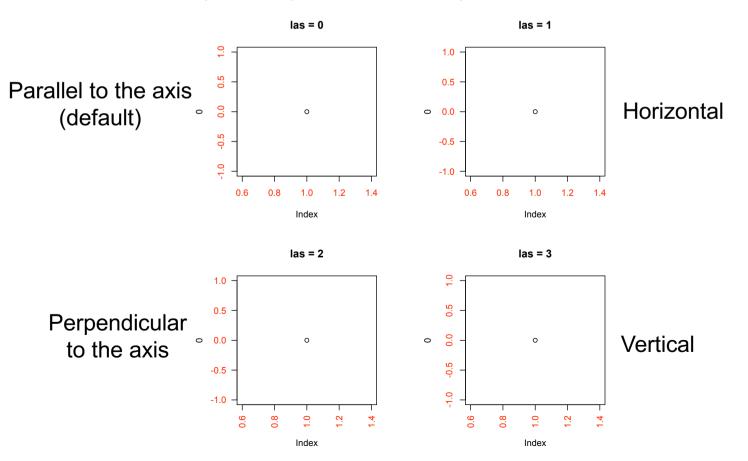
As for col, there are cex.main, cex.sub...





- Position of axis labels : las

par(mfrow = c(2, 2))
for (i in 0:3)
plot(0, las = i, main = paste("las =", i), col.axis = "red")



Functions to interact with graphics

- Reads the position of the graphics cursor when the mouse button is pressed
 - locator()

→ Gives the coordinates of the graphics cursor

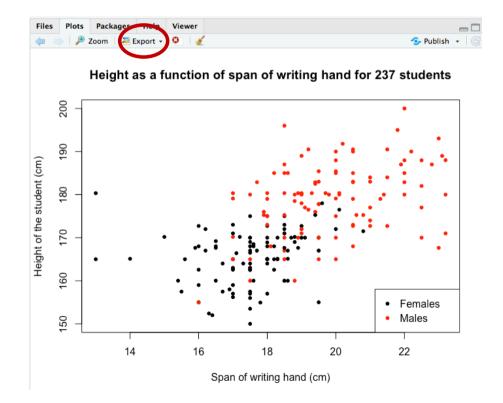
plot(survey\$Wr.Hnd, survey\$Height, pch=20)
locator(2) # 2 points to locate

- identify()

→ Gives the label of the point on the graphics that is closest to the cursor plot(survey\$Wr.Hnd, survey\$Height, pch=20)
 # in order to get the number of the corresponding students : identify(survey\$Wr.Hnd, survey\$Height, rownames(survey))
 # type ESC to quit identify function
 # and get the number of the students corresponding to the points closest to the pointer

Functions linked to graphics file formats

- List of available functions
 - ?Devices
- Main functions
 - pdf(), png(), jpeg(), tiff(), bmp(), postscript(), ...
- Example
 - Save a graphics in a pdf file
 pdf("myfile.pdf")
 #plot instructions
 dev.off() # shut down the current device
- Rstudio Export



Exercise 5 (1/2)

Use the iris dataset available in R > data(iris)

- Choose a graphics file format and save all the graphs you will obtain in this format
- Represent the distribution of the sepal length of all iris with an histogram
 - Choose an appropriate name for the title and the two axes
 - Add above each bar the number of flowers in each class
- Compare sepal length between the different species with a boxplot
 - Choose an appropriate name for the title and the two axes
 - Choose a different colour for each box

Exercise 5 (2/2)

 In the same graphics windows, represent a boxplot for each quantitative variable from the iris dataset allowing to compare the distribution of this variable between the different species

For this purpose :

- Partition the graphics windows
- Write a loop in order to represent one graph for each quantitative variable
- Compare petal and sepal length for all flowers with a scatterplot
 - Choose an appropriate title for each axis
 - Choose a type of points different from the default one
- Represent the same comparison as on previous graphs, with different colours for the different species
 - Add a legend to the graph

Introduction to R software

- R software fundamentals
- Graphics using R
- Basic statistics using R

One dimension descriptive statistics

- Mean : mean
- Median : median
- Variance : var
 - Variance estimation :

$$\hat{\sigma}^2 = \frac{1}{n-1} \sum_{i=1}^n (X_i - \overline{X})^2$$

- Standard deviation : sd
- Quantiles : quantile
- Minimum, maximum : min, max, range
- summary
 - > x=rnorm(1000)
 - > summary(x)

Min. 1st Qu. Median Mean 3rd Qu. Max. -2.89600 -0.64130 0.03306 0.05083 0.71320 3.19600

Exercise 6

For this exercise use the dataset available in humanGenomeSummary.txt file.

- What is the mean and maximum number of protein coding genes per chromosome ?
- Which chromosome contains the highest / smallest number of protein coding genes ?
- Represent the distribution of the number of protein coding genes per chromosome using a bar-chart

Two dimensions descriptive statistics

- Covariance : cov
 - Definition : cov(X,Y) = E(XY) E(X)E(Y)
 - If X and Y are independent then cov(X,Y) = 0
- Correlation : cor

- Definition :
$$R(X,Y) = \frac{\operatorname{cov}(X,Y)}{\sigma(X)\sigma(Y)}$$

Pearson : calculation using observed values of X and Y
 → method = "pearson" (default)

.

- Spearman : calculation using the ranks of observed values of X and Y
 → method = "spearman"
- cor(x,y) : correlation between x and y if x and y are vectors
- cor(m) : correlation between the columns of m if m is a matrix or data.frame

Exercise 7

For this exercise use the dataset available in humanGenomeSummary.txt file.

- Calculate the Pearson correlation coefficient between chromosome size and the number of protein coding genes.
- Calculate the Pearson correlation coefficients between all pairs of quantitative variables. Represent all variables as a function of each other variable from the dataset in order to visualize these correlations (for this purpose use the pairs function).
- Calculate the number of genes, pseudogenes and SNP per kb for each chromosome.
- Are genes and pseudogenes numbers per kb correlated ? And SNP and genes numbers per kb ? To answer these questions calculate the Pearson correlation coefficient and perform a graphical representation.

Hypothesis testing

- Definition
 - Method of statistical inference that allows to accept or reject the validity of hypotheses relative to one or several populations, based on the study of one or several random samples

• Example

– Using the sample of iris available in R, can we say that sepal length is significantly different between *versicolor* and *virginica* species ?

Hypothesis testing principle 1. Choice of the hypothesis to test

- H0 : null hypothesis
 - Formulated in order to be rejected
 - Example
 - H0 = "There is no significant sepal length difference between *versicolor* and *virginica* species"
- H1 : alternative hypothesis
 - Negation of H0
 - Example
 - H1 : "There is a significant sepal length difference between *versicolor* and *virginica* species"
- Hypothesis test
 - Decision rule which allows to decide between H0 and H1

Hypothesis testing principle 1. Choice of the hypothesis to test

- Bilateral test
 - Example
 - Sepal length from *versicolor* species could be either higher or lower than from *viriginica* species
 - In R : alternative = "two.sided" (default)
- Unilateral test
 - The theory predicts the direction of variation
 - Example
 - Is the frequency of lung cancers higher among smokers than among non-smokers ?
 - H0 : The frequency of lung cancers is not significantly higher among smokers than among non-smokers
 - H1 : There is significantly more lung cancers among smokers than among nonsmokers
 - In R : alternative = "less" or "greater" (to specify)

- This choice depend on
 - The type of data
 - The type of hypothesis to be tested
 - The hypotheses that can be done on the characteristics of the studied population (e.g. normality, equality of variances)
- Parametric and non-parametric tests
 - Parametric
 - Assume data come from a type of probability distribution
 - Most often a normal distribution → can be tested using Shapiro-Wilk test : shapiro.test
 - Non-parametric
 - No assumptions about the probability distributions of the variables being assessed
- Paired data
 - Paired e.g. Two treatments have been tested on the same individuals
 - → In R : paired=TRUE
 - Not paired e.g. Two treatments have been tested on different individuals
 - ➔ In R : paired=FALSE by default

A lot of tests implemented in R : apropos("test")

[1] ".valueClassTest"	"Box.test"	"PP.test"	"ansari.t	est" "bartlett.test"	
[6] "binom.test"	"chisq.test"	"cor.test"	"file_test"	"fisher.test"	
[11] "fligner.test"	"friedman.test"	"kruskal.test"	"ks.test"	"mantelhaen.test"	
[16] "mauchley.test"	"mauchly.test"	"mcnemar.test	:" "me	ood.test" "oneway.tes	t"
[21] "pairwise.prop.tes	st" "pairwise.t.test"	"pairwise.wilco	x.test" "po	wer.anova.test" "power.pi	op.test"
[26] "power.t.test"	"prop.test"	"prop.trend.test"	"quade.	test" "shapiro.test"	
[31] "t.test"	"testPlatformEquivale	ence" "testVirtual"	"var.te	st" "wilcox.test"	

- 2 samples comparison
 - Parametric
 - Dispersion
 - Fisher-Snedecor test : var.test
 - Central tendency
 - Student test : t.test(... , var.equal=T)
 - Welch test : t.test
 - Non-parametric
 - Dispersion
 - Ansari test : ansari.test
 - Central tendency
 - Wilcoxon (Mann-Whitney) test : wilcox.test
 - Comparison of distributions
 - Kolmogorov-Smirnov test : ks.test

- Comparison of k samples (k>2)
 - Parametric
 - Dispersion
 - Bartlett test : bartlett.test
 - Central tendency
 - Anova : anova
 - Non-parametric
 - Dispersion
 - Fligner test : fligner.test
 - Central tendency
 - Kruskal-Wallis test (independent samples) : kruskal.test
 - Friedman test (dependant samples) : friedman.test

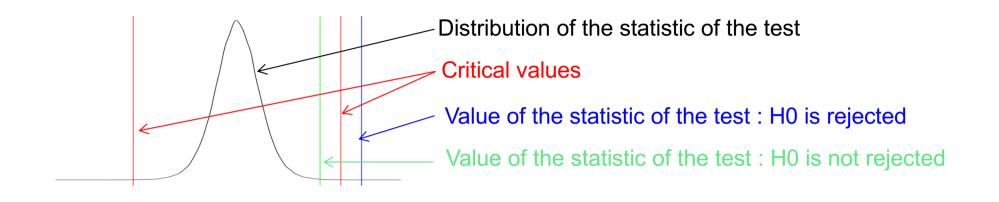
Hypothesis testing principle 3. Choice of a type I error rate

• Different types of errors / risks



- Type I error : α = prob(reject H0 | H0 is true)
 - $-\alpha$ should be chosen by the experimenter before the experiment ... not according to the data !

Hypothesis testing principle 4. Decision

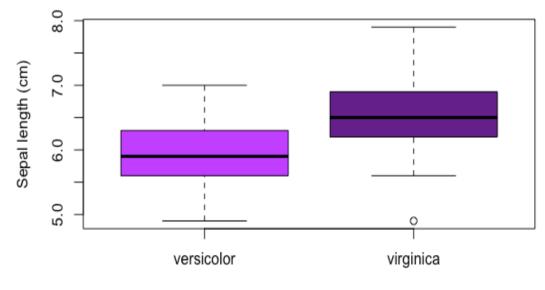


- P-value
 - Probability of obtaining a statistic at least as extreme as the observed value, assuming that the null hypothesis is true
- Decision
 - If p-value < α , we reject H0 with a type I error α
 - If p-value > α , we can not reject H0

• Using the sample of iris available in R, can we say that sepal length is significantly different between *versicolor* and *virginica* species ?

H0 = "There is no significant sepal length difference between *versicolor* and *virginica* species" H1 : "There is a significant sepal length difference between *versicolor* and *virginica* species"

data(iris)
selectspecies = (iris\$Species=="versicolor" | iris\$Species=="virginica")
boxplot(iris\$Sepal.Length[selectspecies] ~ factor(iris\$Species[selectspecies]),
xlab="Species", ylab="Sepal length (cm)", col=c("darkorchid1","darkorchid4"))





Normality of the data ? Shapiro-Wilk test

H0 : we assume this is a sample from a normal distribution # H1 : we do not assume this is a sample from a normal distribution # Let α =0.05

```
shapiro.test(iris$Sepal.Length[iris$Species=="versicolor"])
Shapiro-Wilk normality test
```

```
data: iris$Sepal.Length[iris$Species == "versicolor"]
W = 0.97784, p-value = 0.4647
```

```
shapiro.test(iris$Sepal.Length[iris$Species=="virginica"])
Shapiro-Wilk normality test
```

```
data: iris$Sepal.Length[iris$Species == "virginica"]
W = 0.97118, p-value = 0.2583
```

Conclusion : we can not reject the hypothesis of a normal distribution # We will therefore use parametric tests

Equality of variances ? : Fisher-Snedecor test

H0 : variance of sepal length is not different between *versicolor* and *virginica* species # H1 : this variance is significantly different between *versicolor* and *virginica* species # Let α =0.05

var.test(iris\$Sepal.Length[iris\$Species=="versicolor"], iris\$Sepal.Length[iris\$Species=="virginica"])
#or var.test(iris\$Sepal.Length[selectspecies] ~ factor(iris\$Species[selectspecies]))

F test to compare two variances

Conclusion : We can not reject the hypothesis of variance equality# We will therefore use a Student's t test to compare means

Equality of means ? Student's t test # H0 : mean sepal length is not different between *versicolor* and *virginica* species # H1 : this mean is significantly different between *versicolor* and *virginica* species # Let α =0.05

t.test(iris\$Sepal.Length[iris\$Species=="versicolor"], iris\$Sepal.Length[iris\$Species=="virginica"],
var.equal=TRUE)
#or t.test(iris\$Sepal.Length[selectspecies] ~ factor(iris\$Species[selectspecies]), var.equal=TRUE)

Two Sample t-test

```
data: iris$Sepal.Length[iris$Species == "versicolor"] and iris$Sepal.Length[iris$Species == "virginica"]
t = -5.6292, df = 98, p-value = 1.725e-07
alternative hypothesis: true difference in means is not equal to 0
95 percent confidence interval:
    -0.8818516 -0.4221484
sample estimates:
mean of x mean of y
    5.936    6.588
```

Conclusion : we reject H0, mean sepal length is significantly different between *versicolor* and *virginica* species

Exercise 8

We measured on patients the blood level of a molecule before and after a treatment. These measures are available in molecule.txt file.

Does this treatment allow to significantly increase the level of this molecule ?