

A teal vertical bar is positioned on the left side of the slide, extending from the top to the bottom. A dark blue horizontal line starts from the right edge of this bar and extends across the top of the slide.

# NGS read mapping : answers to questions

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# Exercise 1

## 1. Log file

Proportion of uniquely mapped reads :

Started job on	Feb 04 09:10:08
Started mapping on	Feb 04 09:14:43
Finished on	Feb 04 09:14:54
Mapping speed, Million of reads per hour	327.27
Number of input reads	1000000
Average input read length	50
UNIQUE READS:	
Uniquely mapped reads number	852134
Uniquely mapped reads %	85.24%
Average mapped length	49.84
Number of splices: Total	137459
Number of splices: Annotated (sjdb)	136335
Number of splices: GT/AG	136060
Number of splices: GC/AG	1157
Number of splices: AT/AC	108
Number of splices: Non-canonical	134
Mismatch rate per base, %	0.15%
Deletion rate per base	0.01%
Deletion average length	1.60
Insertion rate per base	0.00%
Insertion average length	1.29
MULTI-MAPPING READS:	
Number of reads mapped to multiple loci	133958
% of reads mapped to multiple loci	13.40%
Number of reads mapped to too many loci	4067
% of reads mapped to too many loci	0.41%
UNMAPPED READS:	
Number of reads unmapped: too many mismatches	0
% of reads unmapped: too many mismatches	0.00%
Number of reads unmapped: too short	7302
% of reads unmapped: too short	0.73%
Number of reads unmapped: other	2239
% of reads unmapped: other	0.22%
CHIMERIC READS:	
Number of chimeric reads	0
% of chimeric reads	0.00%

### History

search datasets

#### RNA-seq data analysis

8 shown

3.33 GB

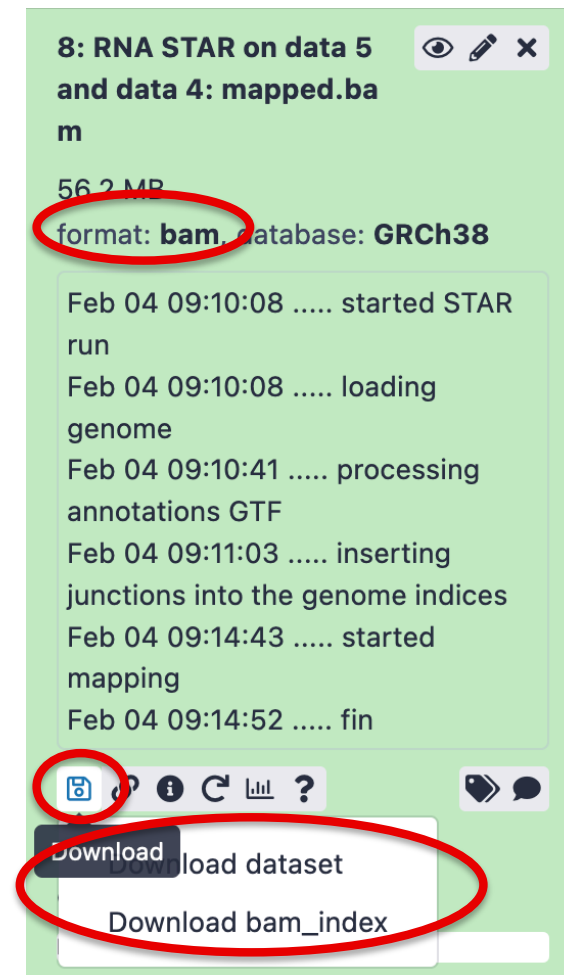
- 8: RNA STAR on data 5 and data 4: mapped.bam
- 7: RNA STAR on data 5 and data 4: splice junctions.bed
- 6: RNA STAR on data 5 and data 4: log
- 5: Homo\_sapiens.GRCh38.105.chr.gtf.gz
- 4: siLuc2\_1000000.fastq.gz
- 3: FastQC on data 1: Raw Data
- 2: FastQC on data 1: Web page

# Exercise 1

## 2. Alignment file

### ■ Galaxy

- STAR provides an alignment in BAM format
- Download this file together with the corresponding index (in the same directory)



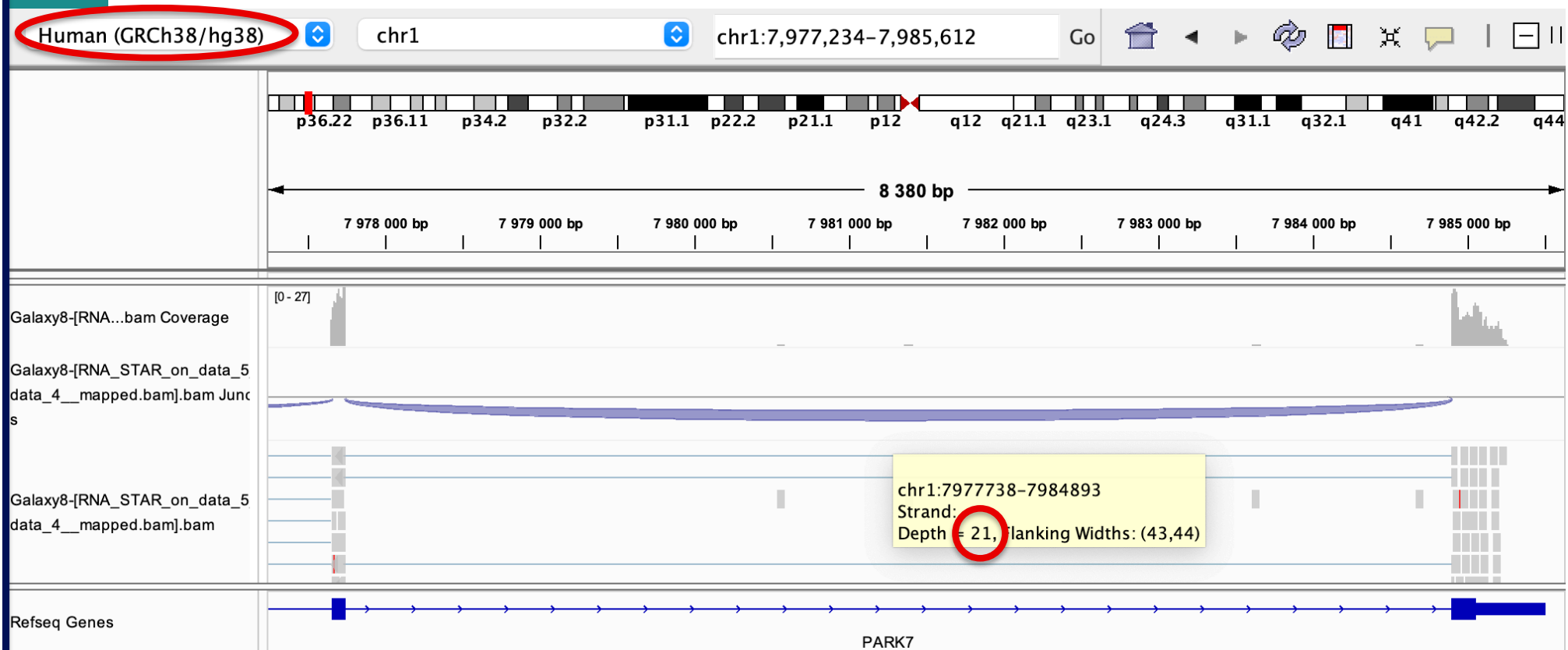
The screenshot shows a Galaxy workflow history entry titled "8: RNA STAR on data 5 and data 4: mapped.bam". The file size is 56.2 MB and the format is "bam" (circled in red). The database is "GRCh38". The workflow steps are: "started STAR run", "loading genome", "processing annotations GTF", "inserting junctions into the genome indices", "started mapping", and "fin". At the bottom, there is a toolbar with icons for download, refresh, and help. A "Download" button is highlighted with a red circle, and a dropdown menu is open showing "load dataset" and "Download bam\_index" (circled in red).

### ■ IGV

- File → Load from file and choose the downloaded BAM file

# Exercise 1

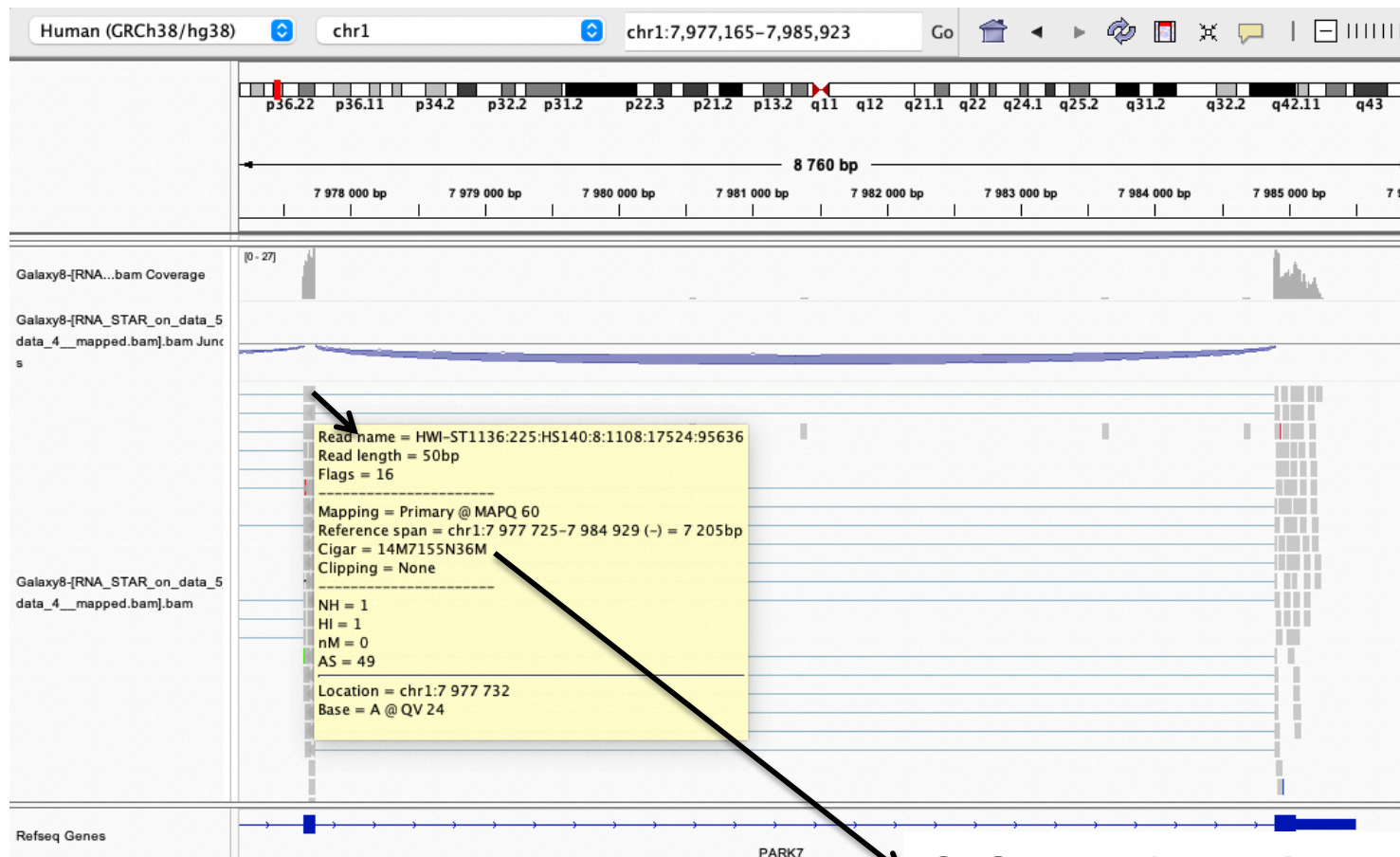
## 2. Splice junction



→ 21 reads span the junction that joins the last 2 exons of *Park7* gene

# Exercise 1

## 2. Splice junction



CIGAR : 14M7155N36M

Intron length :

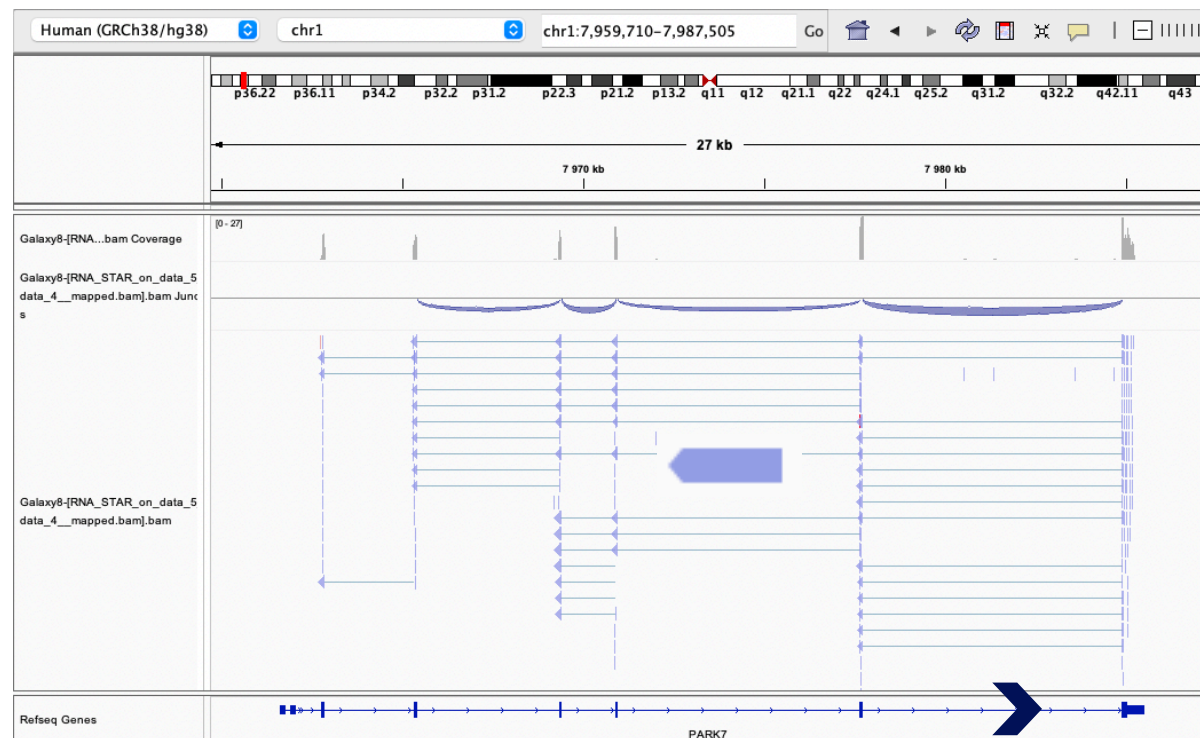
7984893 - 7977738 = 7155

# Exercise 1

## 2. Strand specificity

Right click on BAM file → Color alignments by → read strand

*Park7 :*

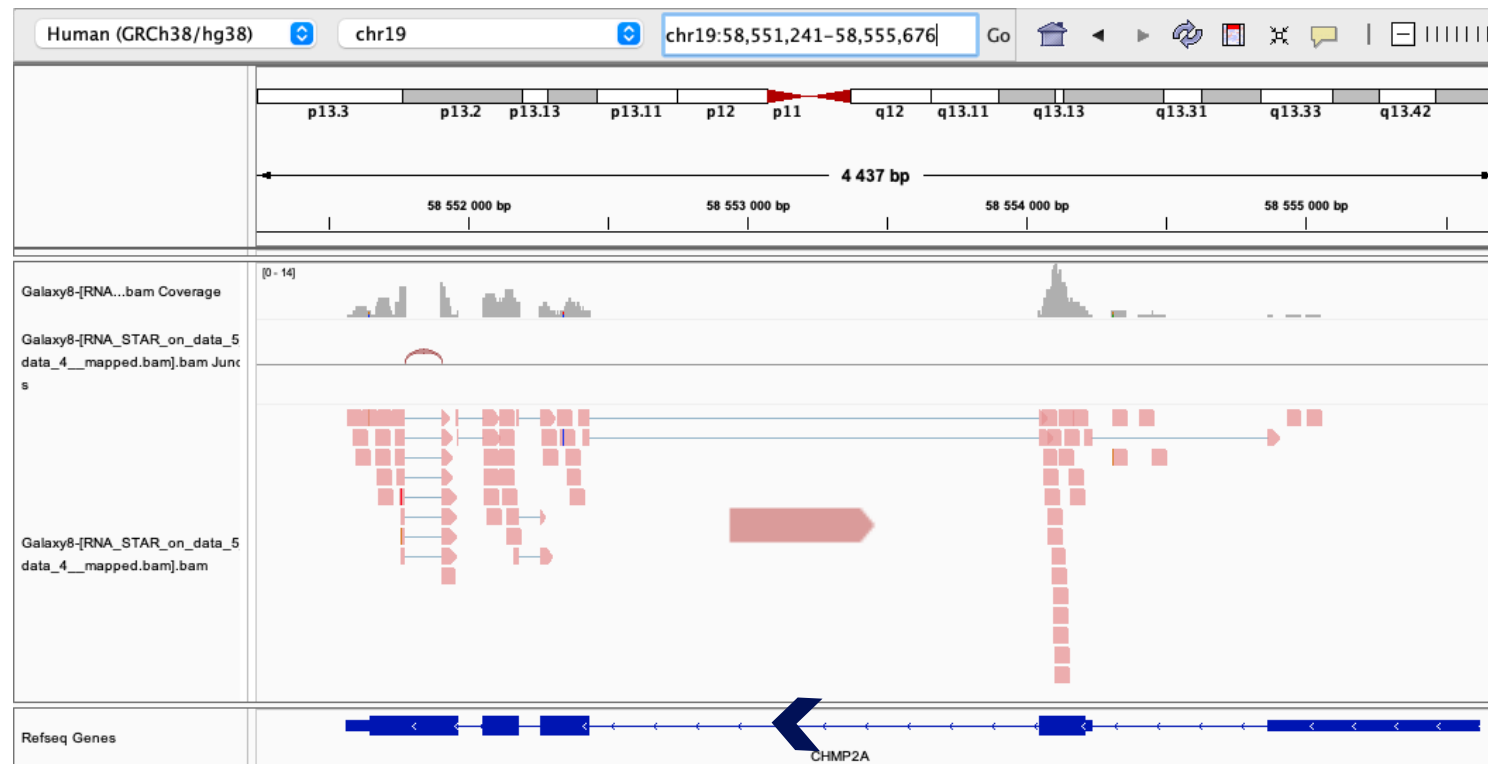


The library has been prepared with a directional mRNAseq protocol which retains strand information :  
reads are in the opposite direction as the transcribed strand

# Exercise 1

## 2. Strand specificity

*Chmp2a* :



The library has been prepared with a directional mRNAseq protocol which retains strand information :  
reads are in the opposite direction as the transcribed strand

# Exercise 1

## 2. Multiple mapped reads

Right click on BAM file → Color alignments by → tag → NH



Number of reported alignments

→ see NH tag in pop-up windows to visualize

color-coding (that can be different from this one) :  1  2  3








































There are multiple aligned reads on this gene



# Exercise 2 - Question 1

## Proportion of uniquely mapped reads

Galaxy : "NGS data analysis training Strasbourg" history

16: RNA STAR on siLuc2_oth er_protocol: mapped.bam	  
15: RNA STAR on siMitf4: ma pped.bam	  
14: RNA STAR on siMitf4: spli ce junctions.bed	  
13: RNA STAR on siMitf4: log	  
12: RNA STAR on siMitf3: ma pped.bam	  
11: RNA STAR on siMitf3: spli ce junctions.bed	  
10: RNA STAR on siMitf3: log	  
9: RNA STAR on siLuc3: map ped.bam	  
8: RNA STAR on siLuc3: spli ce junctions.bed	  
7: RNA STAR on siLuc3: log	  
6: RNA STAR on siLuc2: map ped.bam	  
5: RNA STAR on siLuc2: spli ce junctions.bed	  
4: RNA STAR on siLuc2: log	  

Uniquely mapped reads % | 85.28%

Uniquely mapped reads % | 85.38%

Uniquely mapped reads % | 85.68%

Uniquely mapped reads % | 85.26%

→ This proportion is consistent across samples

# Exercise 2 – Question 2

## *Idh1* gene expression

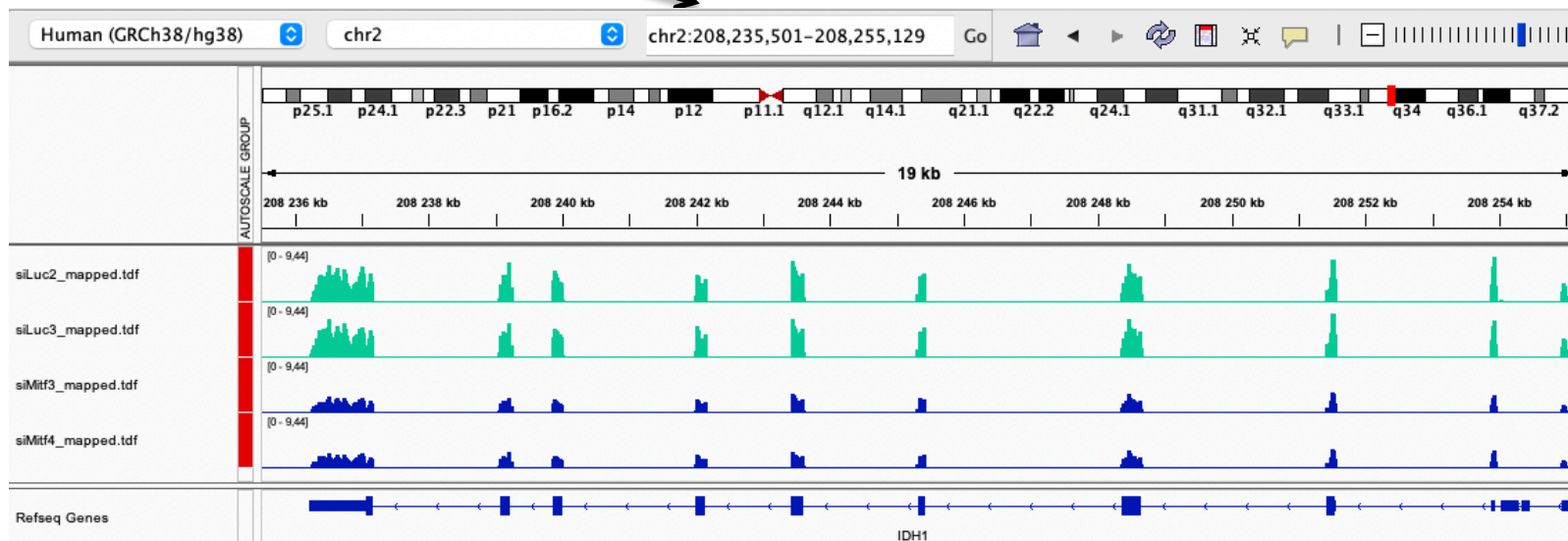
IGV : File → Load from file and select the 4 tdf files

Select all tdf tracks → Right-click → Group Autoscale :

→ IGV automatically adjusts the Y scale to the data range currently in view (this scaling continually adjusts as you move)

→ all tracks are on the same scale

Search for *Idh1*



*Idh1* is under-expressed in siMitf samples compared to siLuc ones

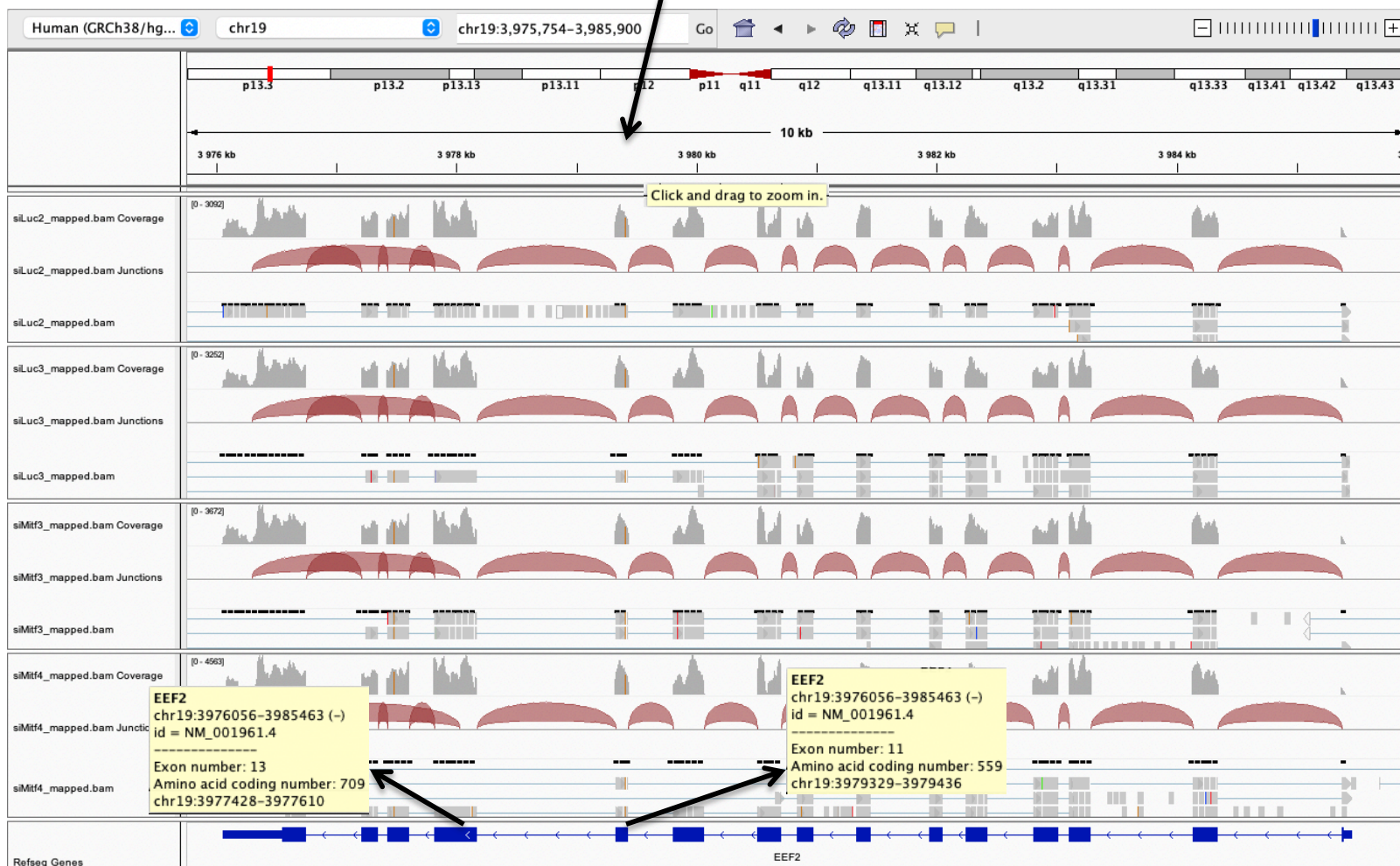
# Exercise 2 – Question 3

- File → new session
- File → load from files and load the 4 BAM files
- Search for *EEF2*



# Exercise 2 – Question 3

Exon numbers are provided on annotation track  
Click and drag on a region to zoom in



# Exercise 2 – Question 3

## ■ *Eef2* exon 11

- chr19:3,979,410 : G in ~100% of the reads, A in the genome



# Exercise 2 – Question 3

## ■ *Eef2* exon 13

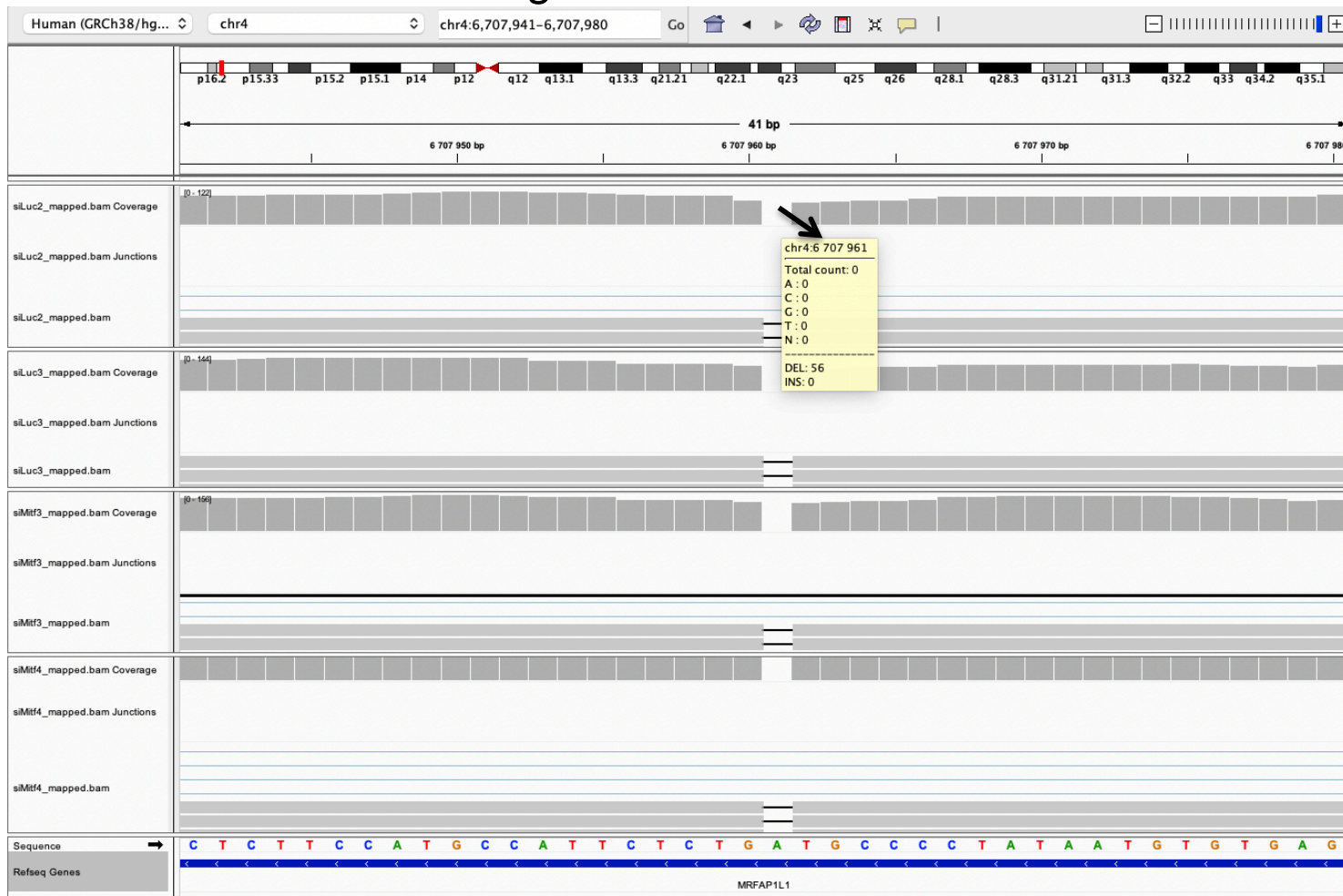
- chr19:3,977,488 : G in ~100% of the reads, A in the genome





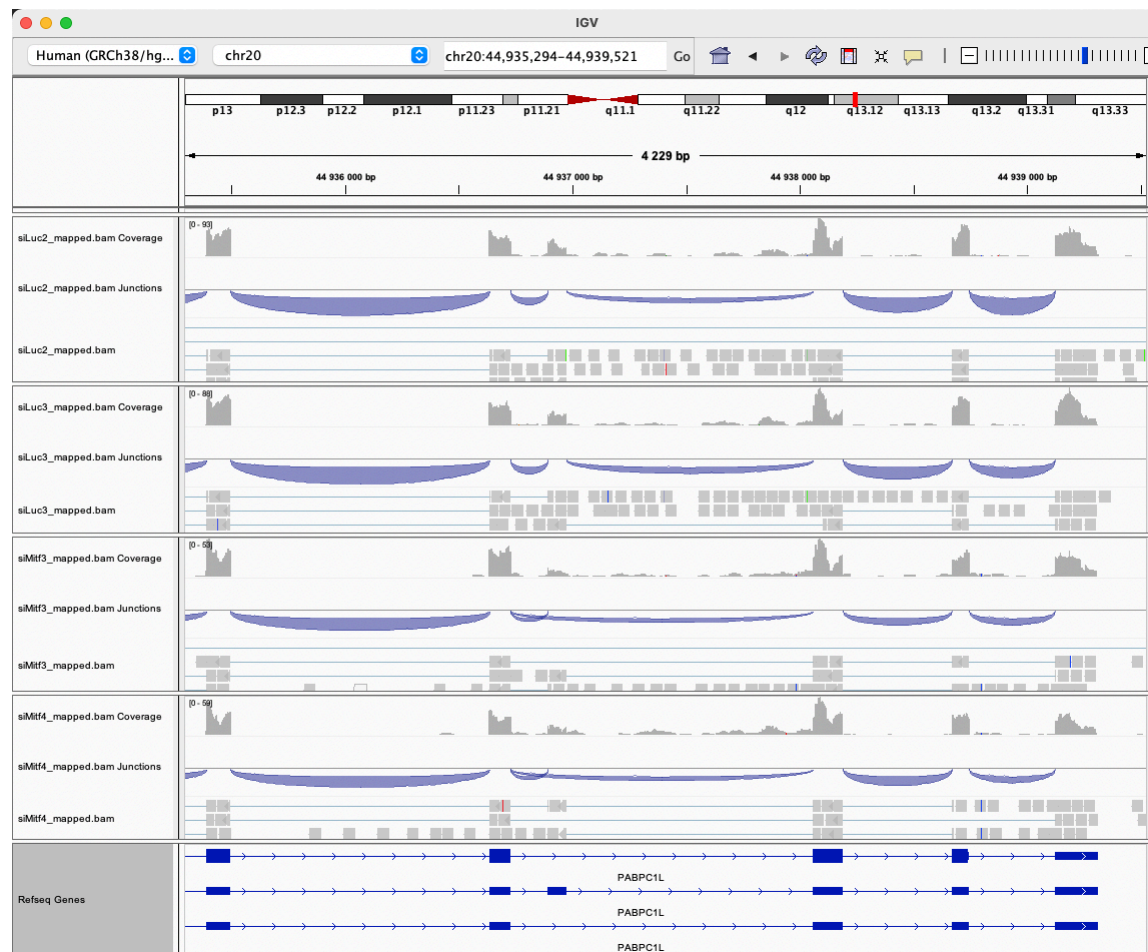
# Exercise 2 – Question 4

- Position chr4:6707961 :
  - Deletion vs reference genome



# Exercise 2 – Question 5

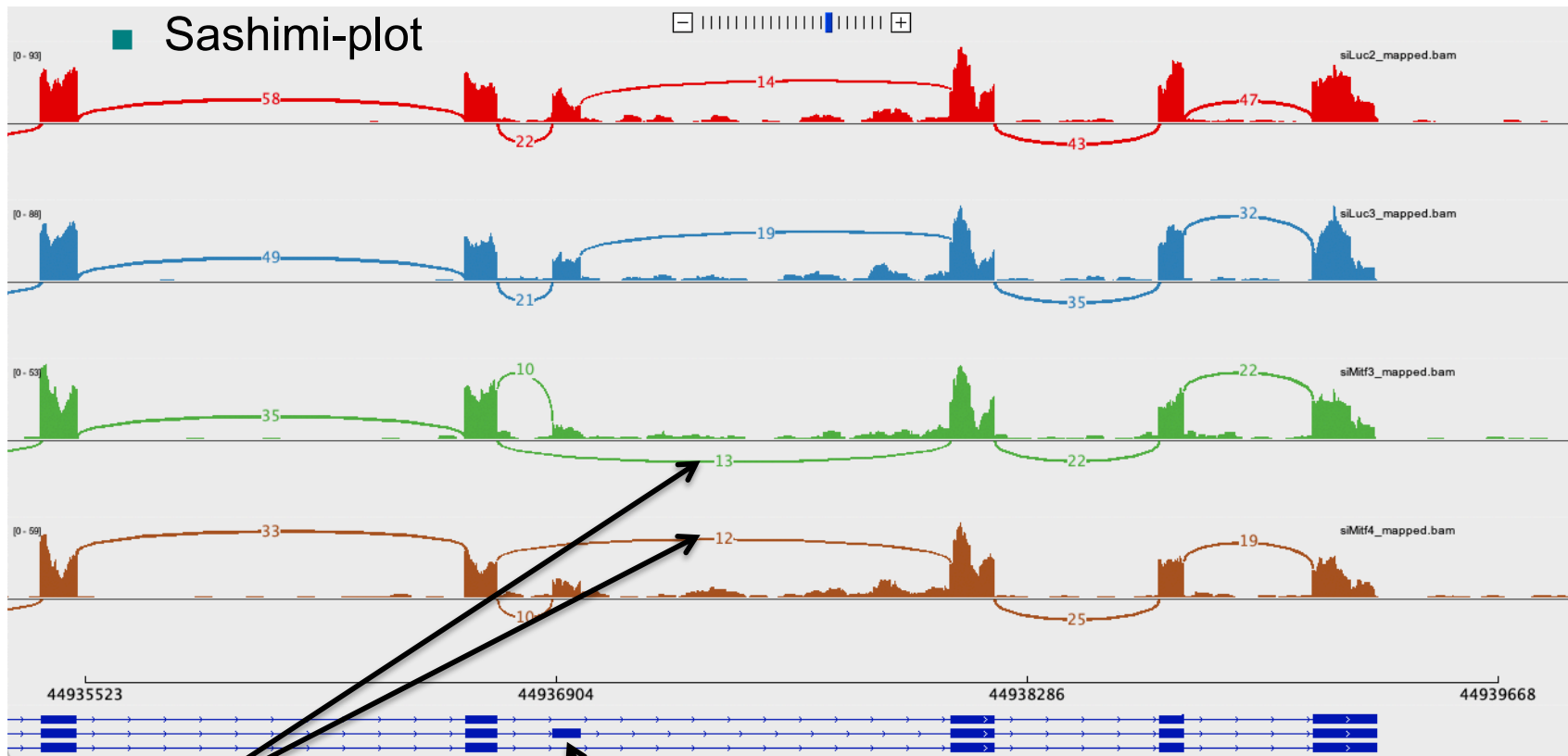
- Region chr20:44,935,294-44,939,521 :
  - Right-click on Refseq Genes track → select Expanded to see all annotated isoforms





# Exercise 2 – Question 5

■ Region chr20:44,935,294-44,939,521 :



We detect an isoform without this exon in siMitf samples

**IGV is only a visualization tool**

**In-depth analysis using paired-end data with more coverage is needed**

# Exercise 2 – Question 5

- If you would like to display Ensembl annotations, you can add this track
  - File → Load from file
  - Select [Homo\\_sapiens.GRCh38.105.chr.sorted.gtf](#) available in [RNAseq/annotations](#) folder



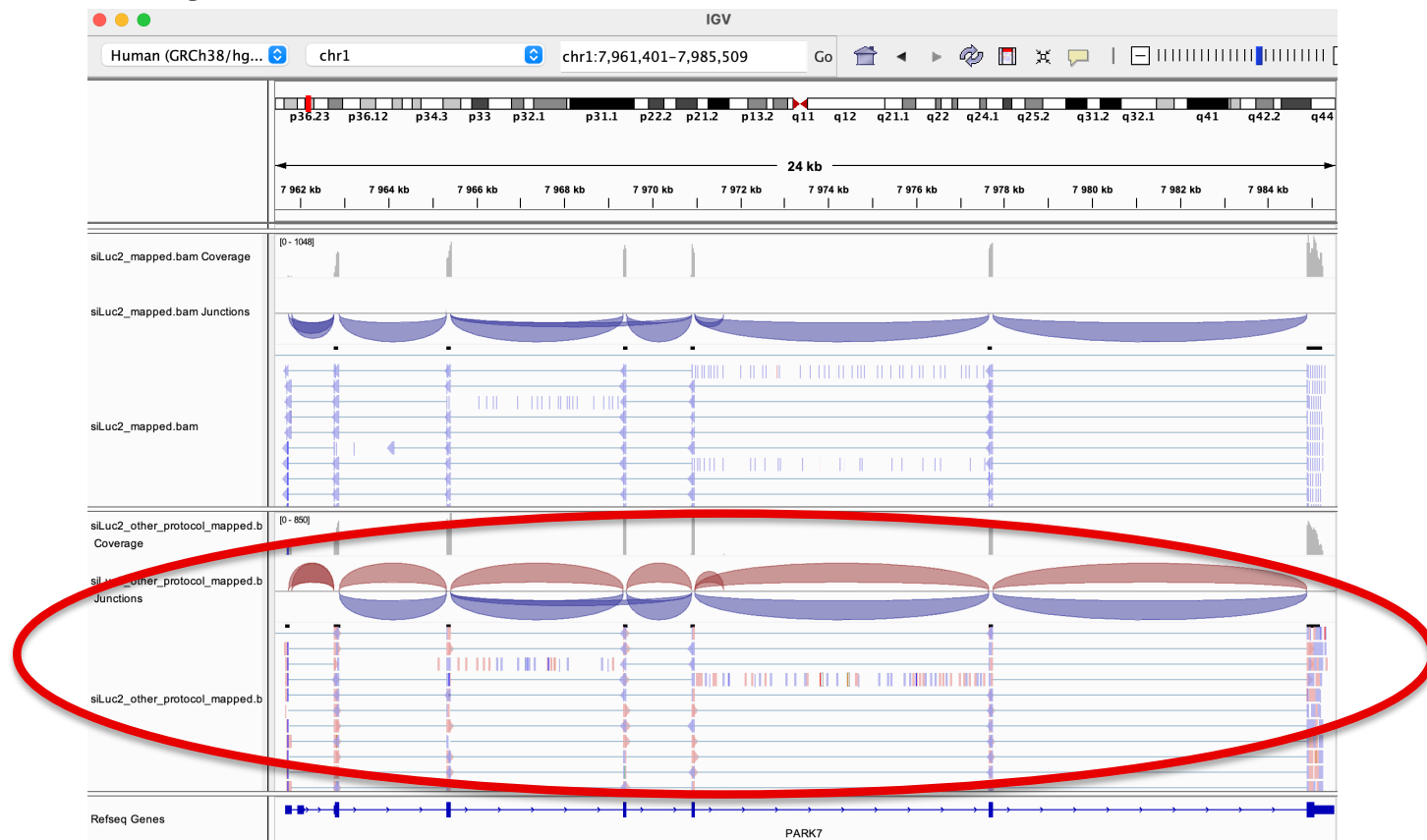
# Exercise 2 – Question 5

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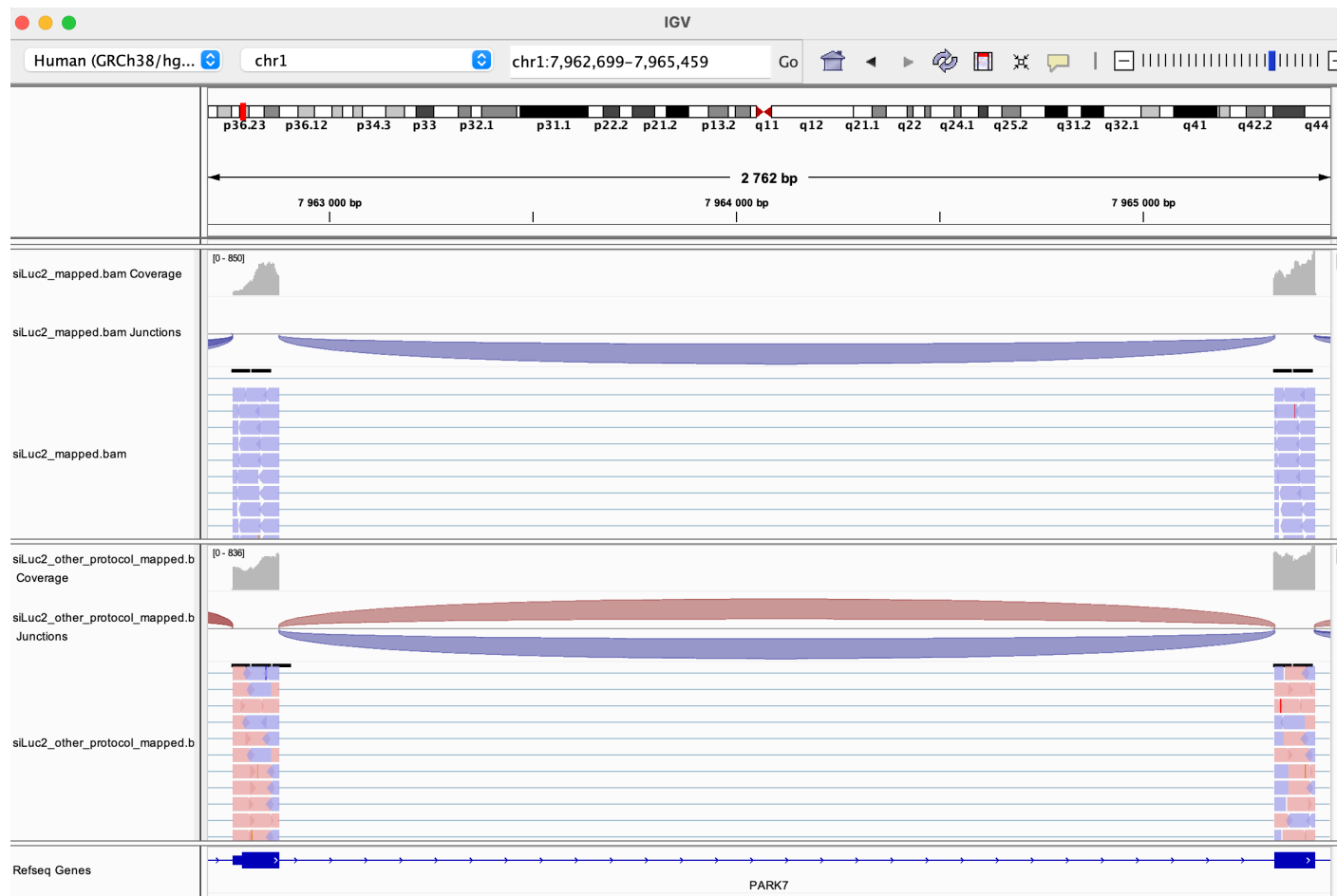
- You can save your IGV session
  - To save the current state of your IGV session to a named session file
  - File → Save Session
  - Data files must stay at the same location
- Use File → Open session to restore a saved session

# Exercise 2 – Question 6

- Remove siLuc3 and siMitf3/4 tracks (Right click on tracks → Remove track)
- File → load from file and select siLuc2\_other\_protocol\_alignment.bam
- Right-click on BAM file → Color alignments by → read strand
- e.g. *Park7* gene



# Exercise 2 – Question 6



This protocol is not directional (it does not preserve strand information)