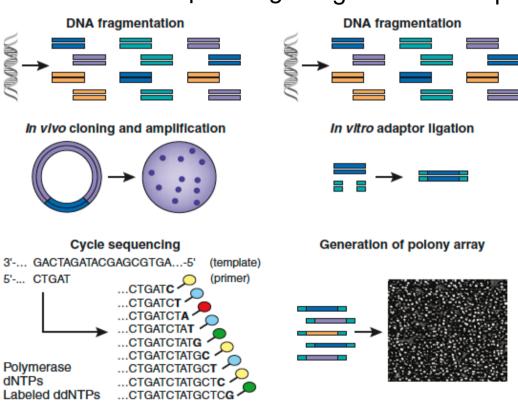
- Introduction to second generation sequencing
- Library preparation
- Amplification
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- Illumina sequencers and throughput
- Other technologies

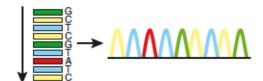
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Conventional vs second generation sequencing

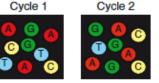
Conventional sequencing 2nd generation sequencing



Electrophorsesis (1 read/capillary)



Cyclic array sequencing (>10⁶ reads/array) Cycle 1 Cycle 2 Cycle 2



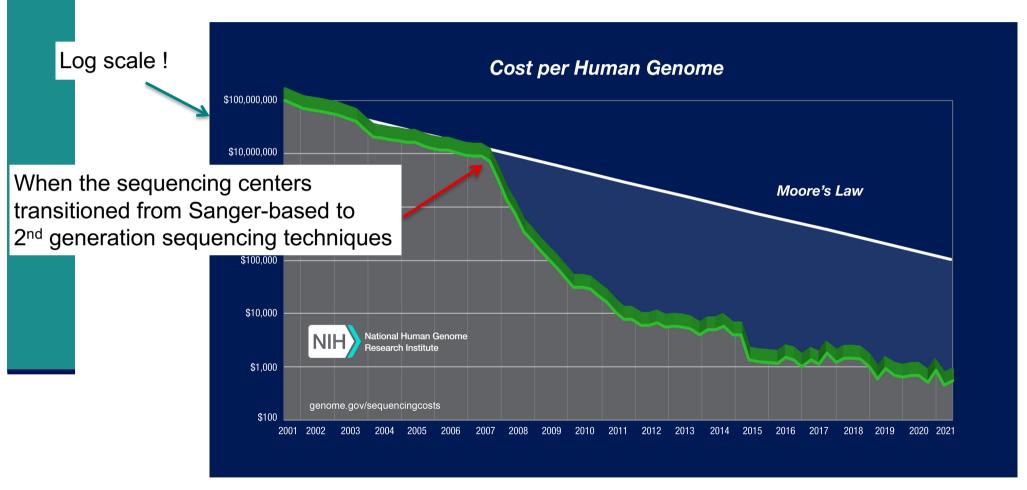


What is base 1? What is base 2? What is base 3?

Three main technologies

- SOLiD DNA Sequencer Applied Biosystems
 - Shendure et al., 2005
- Genome Sequencer FLX+ Roche (formerly 454)
 - Margulies et al., 2005
- Illumina (formerly Solexa)
 - Fedurco et al., 2006
- → Important decrease of
 - Cost per base
 - Time needed to obtain sequences

Decrease of sequencing costs

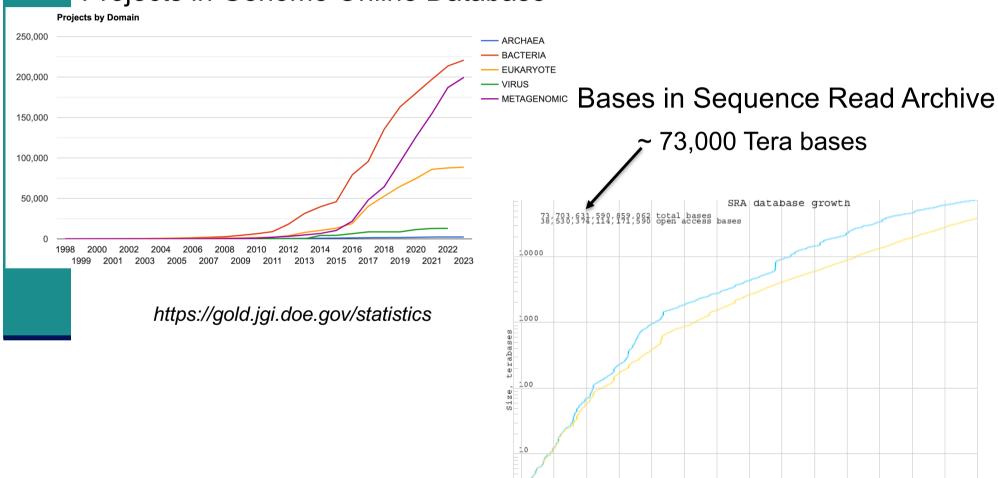


http://www.genome.gov/sequencingcosts/

Human genome with 30X coverage for Illumina sequencing More information on how these costs are calculated: https://www.genome.gov/sequencingcostsdata/

Increase of data volume

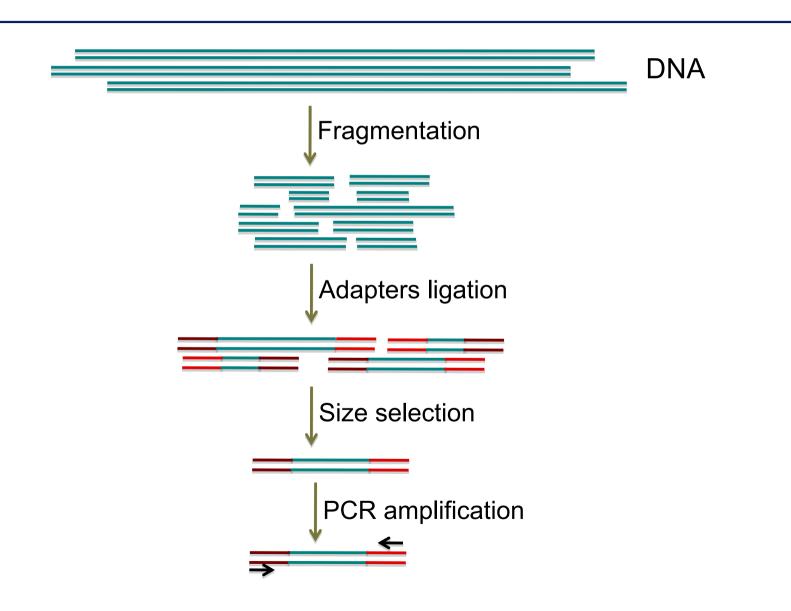
Projects in Genome Online Database



https://www.ncbi.nlm.nih.gov/sra/docs/sragrowth/

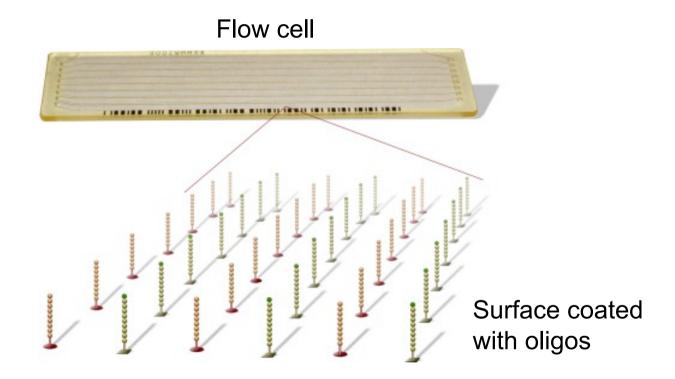
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Library preparation



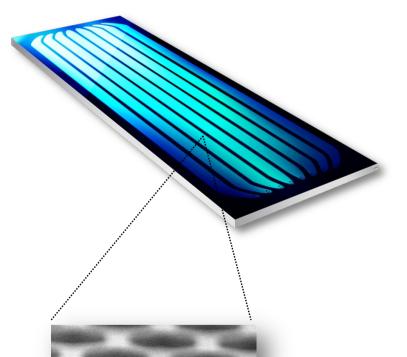
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Fixation of DNA fragments on a solid support



Fixation of DNA fragments on a solid support

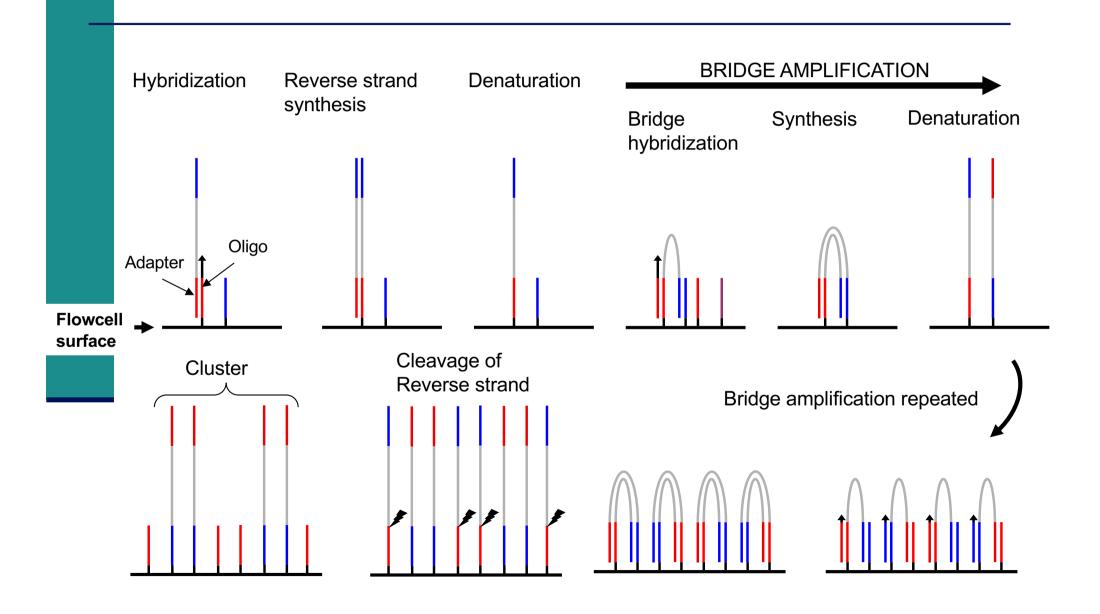
Patterned Flow Cell: billions of ordered wells



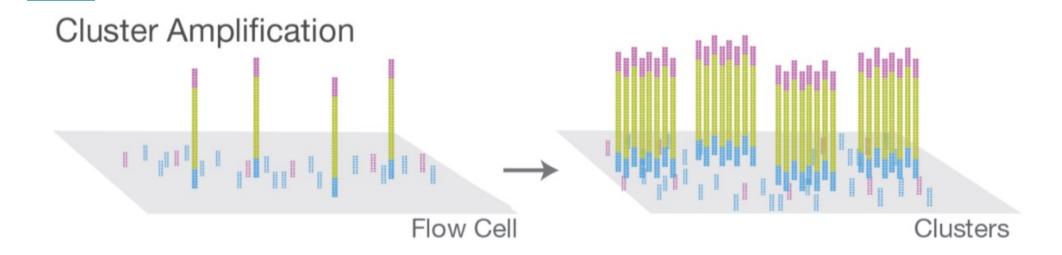
- → More reads
- → Faster run time

iSeq100, NextSeq1000/2000, Hiseq3000/4000/X, NovaSeq6000/X sequencers

Amplification: method

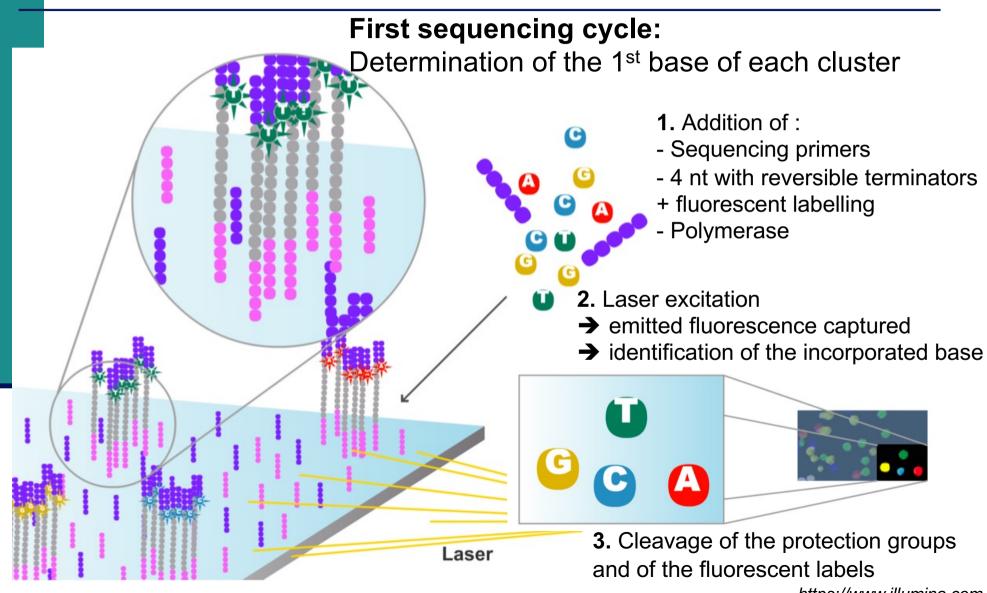


Amplification: result



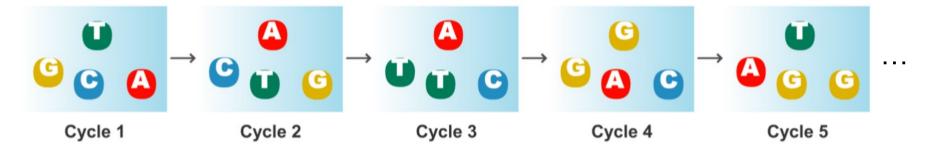
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Sequencing by synthesis



Sequencing by synthesis

As many sequencing cycles as the number of bases needed in the resulting read



eg cluster 1 = TAAGT...

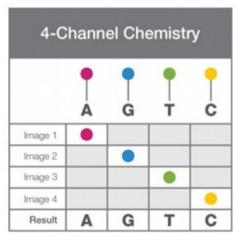
→ 1 read

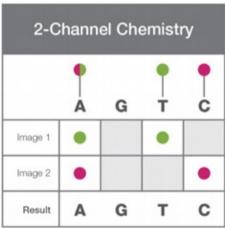
Each lane (top and bottom) is divided into swaths composed of tiles

→ 1 image per tile per color, for each cycle

2-channel sequencing by synthesis

- Evolution of the original 4-channel SBS technology
 - 2 fluorescent dyes
 - Only 2 images per cycle instead of 4
 - Accelerates sequencing and data processing times
- Clusters seen in red images → C
- Clusters seen in green images → T
- Clusters observed in both red and green images → A
- Unlabelled clusters → G

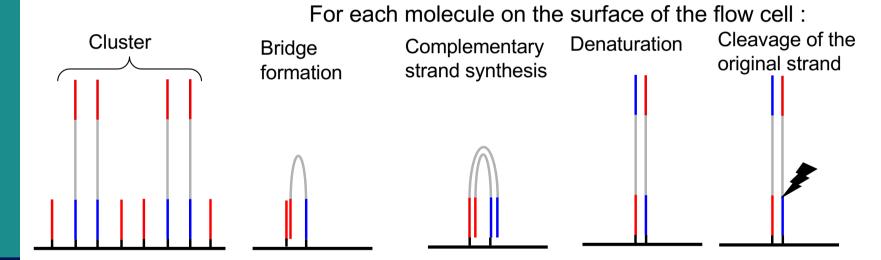




Paired-end sequencing



- Sequencing of read 1: previously described method
- Then sequencing of read 2 :



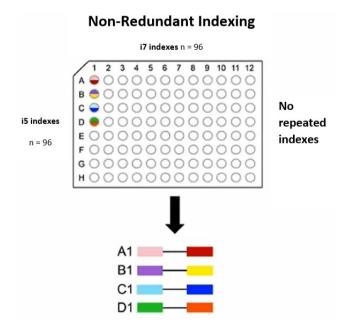
- → Sequence the other end of the original molecule
- → Step performed on the flow cell in the sequencer: keep the position of clusters This information allows to link pairs of sequences

Multiplexing

- Add a barcode (index) specific to each sample
- Sequencing of several samples together
- Single indexing : 1 barcode

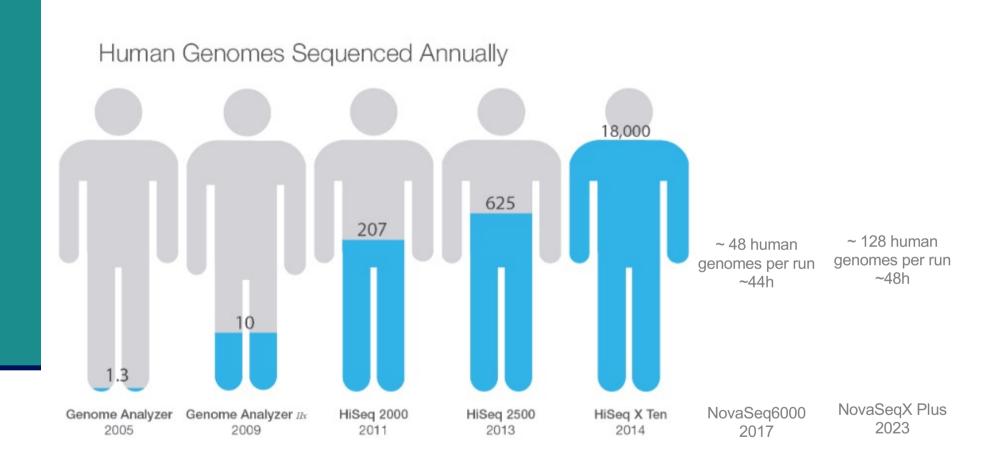


- Dual indexing : 2 barcodes
 - Unique dual indexes : distinct index sequences for each of the two indexes



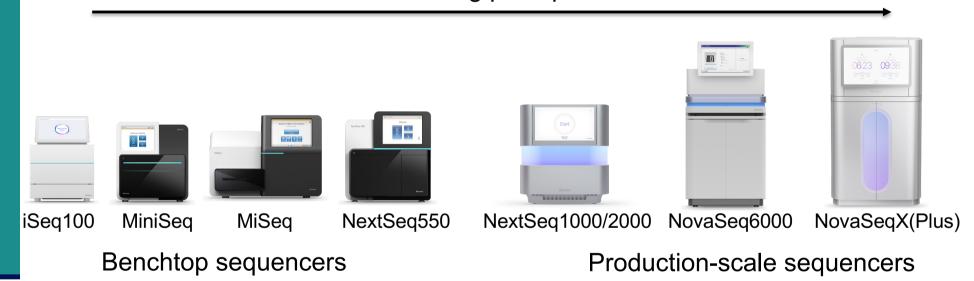
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Illumina throughput over time

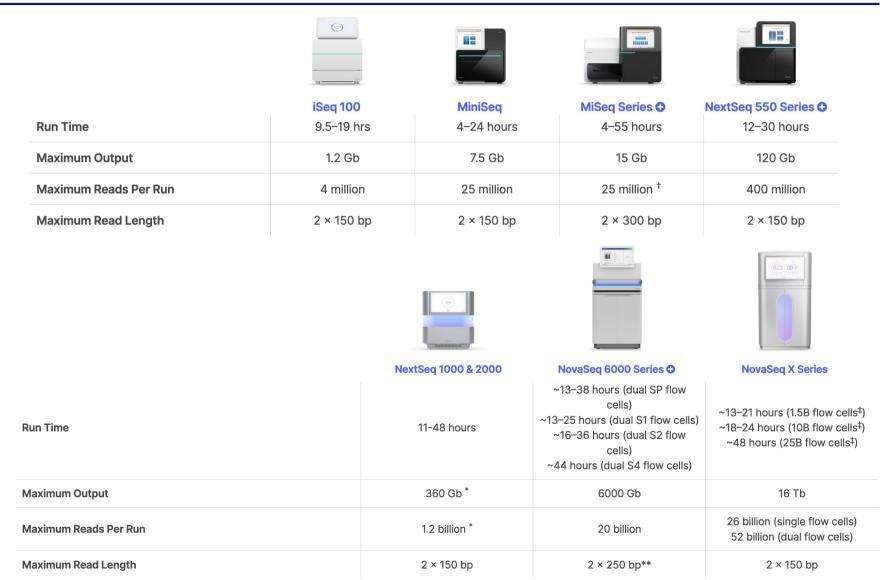


Illumina sequencers

Increasing system price and output Decreasing price per Gb



Illumina sequencers



Illumina NextSeq2000

■ 3 types of flow cell

Flow cells*	Maximum number of clusters/flow cells#		Read length options (in bp)		
P1	100 M				300
P2	400 M		100	200	300
P3	1000 M	50	100	200	300

^{*} Each type of flow cell can be used indiscriminately for single read (SR) or paired-end (PE) sequencing: e.g. a 100 bp kit can be used as SR100, PE50, PE25x75 ...

^{# 100} M clusters = 100 M reads in SR and 2x100 M in PE sequencing

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Other technologies

- Short reads
 - ~ up to 300 bases
 - Genome resequencing, expression and epigenetic analysis
 - MGI, Ultima Genomics, Pacific Biosciences, Element Biosciences, ...
- Long reads
 - ~ several kb
 - De novo sequencing, structural variation analysis, isoform sequencing
 - Oxford Nanopore Technologies, Pacific Biosciences
 - Longer reads, but lower throughput